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(54) METHODE DE REDUCTION DE SUBSTANCES ODORANTES
(54) METHOD OF REDUCING BOAR TAINT

(57)

A method of preventing or reducing boar taint is described. The method involves increasing the metabolism of skatole in a pig by modulating the activity or expression of the enzymes involved in skatole metabolism.

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(54) **METHODE DE REDUCTION DE SUBSTANCES ODORANTES**
(54) **METHOD OF REDUCING BOAR TAINT**

(57) A method of preventing or reducing boar taint is described. The method involves increasing the metabolism of skatole in a pig by modulating the activity or expression of the enzymes involved in skatole metabolism.

ABSTRACT OF THE DISCLOSURE

A method of preventing or reducing boar taint is described. The method involves increasing the metabolism of skatole in a pig by modulating the activity or expression of the enzymes involved in skatole metabolism.

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Title: Method of Reducing Boar Taint

FIELD OF THE INVENTION

The invention relates to a method for preventing boar taint by decreasing the levels of skatole in fat. Levels of skatole in fat can be decreased by enhancing the metabolism and clearance of skatole in pigs.

BACKGROUND OF THE INVENTION

Male pigs that are raised for meat production are usually castrated shortly after birth to prevent the development of off-odors and off flavors (boar taint) in the carcass. Boar taint is primarily due to high levels of either the 16-androstene steroids (especially 5α (-androst-16-en-3-one) or skatole in the fat. Recent results of the EU research program AIR 3 - PL94 -2482 suggest that skatole contributes more to boar taint than androstenone (Bonneau, M., 1997).

Skatole is produced by bacteria in the hindgut which degrade tryptophan that is available from undigested feed or from the turnover of cells lining the gut of the pig (Jensen and Jensen, 1995). Skatole is absorbed from the gut and metabolised primarily in the liver (Jensen and Jensen, 1995). High levels of skatole can accumulate in the fat, particularly in male pig, and the presence of a recessive gene Ska^1, which results in decreased metabolism and clearance of skatole has been proposed (Lundström et al., 1994; Friis, 1995). Skatole metabolism has been studied extensively in ruminants (Smith, et al., 1993), where it can be produced in large amounts by ruminal bacteria and results in toxic effects on the lungs (reviewed in Yost, 1989). The metabolic pathways involving skatole have not been well described in pigs. In particular, the reasons why only some intact male pigs have high concentrations of skatole in the fat are not clear. Environmental and dietary factors are important (Kjeldsen, 1993; Hansen et al., 1995) but do not sufficiently explain the reasons for the variation in fat skatole concentrations in pigs. Claus et al. (1994) proposed high fat skatole concentrations are a result of an increased intestinal skatole production due to the action of androgens

and glucocorticoids. Lundström et al. (1994) reported a genetic influence on the concentrations of skatole in the fat, which may be due to the genetic control of the enzymatic clearance of skatole. The liver is the primary site of metabolism of skatole and liver enzymatic activities could be the controlling factor of skatole deposition in the fat. Bæk et al.(1995) described several liver metabolites of skatole found in blood and urine with the major being MII and MIII. MII, which is a sulfate conjugate of 6-hydroxyskatole (pro-MII), was only found in high concentrations in plasma of pigs which were able to rapidly clear skatole from the body, whereas high MIII concentrations were related to slow clearance of skatole. Thus the capability of synthesis of MII could be a major step in a rapid metabolic clearance of skatole resulting in low concentrations of skatole in fat and consequently low levels of boar taint.

In view of the foregoing, further work is needed to fully understand the metabolism of skatole in pig liver and to identify the key enzymes involved. Understanding the biochemical events involved in skatole metabolism can lead to novel strategies for treating, reducing or preventing boar taint. In addition, polymorphisms in these candidate genes may be useful as possible markers for low boar taint pigs.

SUMMARY OF THE INVENTION

Broadly stated, the present invention relates to a method for reducing boar taint comprising enhancing or increasing the metabolism of skatole in a pig.

The metabolism of skatole in pigs involves Phase I oxidation reactions carried out by cytochrome P450, in particular CYP2E1, and Phase II conjugation reactions carried out by glucuronyl transferases, sulfotransferases and glutathionine transferases.

With regard to the Phase I reactions, the inventors have shown that decreased expression of CYP2E1 in the livers of pigs is correlated with high levels of skatole in the fat. The inventors have also shown that synthesis of metabolite F-1 (indole-3-carbinol) is correlated with low levels of skatole in the fat. Accordingly, in one aspect, the present invention provides a method for reducing boar taint comprising

enhancing the activity of CYP2E1 in a pig. The activity of CYP2E1 can be enhanced by using substances which (a) increase the activity of CYP2E1; or (b) induce or increase the expression of the CYP2E1 gene.

With regard to the Phase II reactions, the inventors have shown that the formation and sulfation of the metabolite 6-hydroxy-3-methylindole (pro-MII) is related to low fat skatole levels while high rates of glucuronidation were correlated with high skatole levels in fat. Accordingly, in another aspect, the present invention provides a method for reducing boar taint comprising enhancing the activity of a sulfotransferase in a pig. In a further aspect, the present invention provides, a method for reducing boar taint comprising inhibiting the activity of a glucuronyl transferase in a pig.

The present invention also includes a method of screening for a substance that regulates skatole metabolism in a pig. In one embodiment, the present invention provides a method for screening a substance that activates CYP2E1 activity or induces transcription and/or translation of the gene encoding CYP2E1. In another embodiment, the present invention provides a method for screening for a substance that enhances the activity of a sulfotransferase or enhances the transcription or translation of a gene encoding a sulfotransferase in a pig. In a further embodiment, the present invention includes a method for screening for a substance that inhibits glucuronidation activity or inhibits the transcription or translation of a gene encoding a glucuronyl transferase in a pig.

The present invention also includes a pharmaceutical composition for use in treating boar taint comprising an effective amount of a substance which regulates skatole metabolism in a pig and/or a pharmaceutical acceptable carrier, diluent or excipient.

The present invention further includes a method for producing pigs that have a lower incidence of boar taint comprising selecting pigs that express high levels of CYP2E1 and/or sulfotransferase and/or low levels of glucuronyl transferase; and breeding the selected pigs.

Other objects, features and advantages of the present invention will become apparent from the following detailed description. It should be understood, however, that the detailed description and the specific examples while indicating preferred embodiments of the invention are given by way of illustration only, since various changes and modifications within the spirit and scope of the invention will become apparent to those skilled in the art from this detailed description.

BRIEF DESCRIPTION OF THE DRAWINGS

The invention will now be described in relation to the drawings in which:

Figure 1 is a HPLC chromatogram of skatole metabolites produced by liver microsomes as detected by UV absorbance, (A) and fluorescence, (B). Peaks UV-2 and F-4 were identified as MIII and pro-MII, respectively; IS, internal standard (indole-3-acetonitrile); SK, skatole; UV-1, 3-methyloxindole and indole-3-carbinol to F-3 were not identified.

Figure 2 is the formation of skatole metabolites in liver microsomes. Metabolite F-1 is not presented here because of its much larger peak area than that of the other metabolites. Its synthesis pattern over time followed that of the other metabolites. Each data point represents the mean of duplicate assays performed for 2 pigs.

Figure 3 shows the effect of SKF 525A on formation of skatole metabolites in liver microsomes. Each data point represents the mean of duplicate assays performed for 3 pigs.

Figure 4 shows the effect of metyrapone on formation of skatole metabolites in liver microsomes. MIII could not be separated by HPLC from an unidentified metabolite of metyrapone, and therefore it was not possible to quantify MIII formation. Each data point represents the mean of duplicate assays performed for 3 pigs.

Figure 5 shows the effect of DAS on formation of skatole metabolites in liver microsomes. Each data point represents the mean of duplicate assays performed for 3 pigs.

Figure 6 shows the effect of chlorzoxazone on formation of skatole metabolites in liver microsomes. Each data point represents the mean of duplicate assays performed for 3 pigs.

Figure 7 shows the glucuronidation of liver microsomal metabolites of skatole. Each data point represents the mean of duplicate assays performed for 3 pigs. UDPGA is uridine 5'-diphosphoglucuronyl. **Differs from control, $P < .01$; ***differs from control, $P < .001$.

Figure 8 shows the sulfation of liver microsomal metabolites of skatole. The mean concentrations of skatole metabolite F-2 compared to controls were: 30 min, 301%; 60 min, 890%; 90 min, 964%, and did not differ from controls ($P = .274$). Each data point represents the mean of duplicate assays performed for 3 pigs. PAPS is adenosine 3'-phosphate 5'-phosphosulfate. *Differs from control, $P < .05$; **differs from control, $P < .01$.

Figure 9 shows the HPLC chromatogram of glucuronidation (A) and sulfation (B) of pro-MII (6-hydroxy-3-methylindole) in liver microsomes. Peaks 1 and 2, products of pro-MII glucuronidation; peak 3, product of pro-MII sulfation; IS, internal standard (indole-3-acetonitrile). The metabolites were detected by fluorescence, excitation wavelength, 285 nm, and emission wavelength, 340 nm.

DETAILED DESCRIPTION OF THE INVENTION

1. Methods of Enhancing Skatole Metabolism

As hereinbefore mentioned, the present invention relates to a method for preventing boar taint by enhancing the metabolism of skatole in a pig. The metabolism of skatole can be enhanced by altering the Phase I oxidation reactions carried out by cytochrome P450 and/or altering the Phase II conjugation reactions carried out by glucuronyl transferase and sulfotransferases.

Phase I Metabolism

In Phase I of skatole metabolism, the cytochrome P450 enzyme CYP2E1 (or P450IIE1) catalyzes the biotransformation of skatole to 6-hydroxy-skatole (pro-MII) and other metabolites including F-1, an

indole-3-carbinol. The inventors have shown that decreased expression of CYP2E1 in the liver of pigs (measured by Western analysis) is correlated with high skatole levels in the fat. The inventors have demonstrated that general inhibitors of cytochrome P450 and specific inhibitors of CYP2E1 reduced the production of skatole metabolites by liver microsomes.

Accordingly, the present invention provides a method for reducing or preventing boar taint comprising enhancing the activity of CYP2E1 in a pig. The activity of the CYP2E1 enzyme can be enhanced by administering a substance (a) that increases the activity of the CYP2E1 enzyme; or (b) a substance that induces or increases the expression of the CYP2E1 gene. Substances that increase the activity of the CYP2E1 enzyme or induce or increase the expression of the CYP2E1 gene include substances that stabilize the protein and/or mRNA such as ethanol and acetone. CYP2E1 can also be induced by nutritional related conditions such as fasting and diabetes. The activity of the CYP2E1 may also be enhanced using gene therapy whereby a nucleic acid sequence encoding a CYP2E1 enzyme is introduced into a pig, either *ex-vivo* or *in vivo*. A nucleic acid sequence encoding a CYP2E1 enzyme may be obtained from GenBank Accession No. AB000885 or Swiss-Prot:P79383.

The inventors have also shown that the production of the skatole metabolites F-1 (an indole-3-carbinol) and pro-MII (6-hydroxyskatole) were negatively correlated to skatole levels in fat.

Accordingly, the present invention also provides a method for detecting the capacity for boar taint comprising determining the production of F1 and/or pro-MII in a sample from a pig.

Phase II Metabolism

In Phase II of skatole metabolism, skatole and its metabolites (produced in Phase I) are conjugated by glucuronidation and sulfation in the liver. The inventors have shown that the formation and subsequent sulfation of the metabolite 6-hydroxyskatole (pro-MII) to 6-sulfatoxyskatole (MII) was related to low fat skatole levels, while high rates of glucuronidation were correlated with high skatole levels in fat.

This suggests that high sulfotransferase and low glucuronyl transferase activity in the liver is necessary for effective clearance of skatole by the pig.

Accordingly, in another aspect, the present invention provides a method for reducing boar taint comprising enhancing the activity of a sulfotransferase in a pig. The activity of a sulfotransferase can be enhanced by administering a substance and substance (a) that increases the activity of a sulfotransferase enzyme; or (b) a substance that induces or increases the expression of a sulfotransferase gene. In another embodiment, the activity of the sulfotransferase enzyme can also be enhanced by administering a nucleic acid sequence encoding a sulfotransferase enzyme into a pig, either *ex vivo* or *in vivo*.

Nucleic acid sequences encoding a sulfotransferases may be obtained from known sources. For example, the sequence of a human sulfotransferase may be obtained from GenBank Accession No. U52852, and the sequence of a guinea pig sulfotransferase may be obtained from GenBank accession on number L11117. Two families of sulfotransferases have been described in animals, the phenol sulfotransferase and the hydroxysteroid sulfotransferase. It is likely that the phenol sulfotransferase is responsible for the Phase II metabolism of skatole in pigs. Five sulfotransferases have been isolated and cloned from human (Weishilboum et al., 1997). Three subfamilies of sulfotransferases exist in rats. Sulfotransferases have also been characterized in mice, guinea pigs, bovines as well as in plants (Matsui and Homma, 1994; Runge-Morris, 1997). Recently, the x-ray crystal structure of sulfotransferase has been reported (Negishi et al. 1998). Estrogens decrease while androgens increase sulfotransferase activity in rats (Matsui and Homma, 1994), while there are no pronounced differences in sulfotransferase between sexes in humans (Tamellini et al., 1991). Genetic polymorphisms for sulfotransferase have been reported for several mammalian species (Weishilboum et al., 1997) and may also exist in the pig. The significant interindividual variability (Pacifici et al., 1994) and genetic

polymorphism of sulfotransferases likely explains why some pigs are more susceptible to developing boar taint.

In a further aspect, the present invention provides, a method for reducing boar taint comprising inhibiting the activity of a glucuronyl transferase in a pig. The activity of a glucuronyl transferase enzyme may be inhibited by administering a substance (a) that inhibits the activity of the glucuronyl transferase enzyme; or (b) a substance that inhibits the expression of the glucuronyl transferase gene. Substances that inhibit the activity of a glucuronyl transferase enzyme include antibodies to the enzyme. Antibodies capable of binding to a glucuronyl transferase enzyme can be prepared using techniques known in the art. For example, by using a peptide or the entire enzyme, polyclonal antisera or monoclonal antibodies can be made using standard methods. A mammal, (e.g., a mouse, hamster, or rabbit) can be immunized with an immunogenic form of the peptide which elicits an antibody response in the mammal. Techniques for conferring immunogenicity on a peptide include conjugation to carriers or other techniques well known in the art. For example, the peptide can be administered in the presence of adjuvant. The progress of immunization can be monitored by detection of antibody titers in plasma or serum. Standard ELISA or other immunoassay procedures can be used with the immunogen as antigen to assess the levels of antibodies. Following immunization, antisera can be obtained and, if desired, polyclonal antibodies isolated from the sera.

To produce monoclonal antibodies, antibody producing cells (lymphocytes) can be harvested from an immunized animal and fused with myeloma cells by standard somatic cell fusion procedures thus immortalizing these cells and yielding hybridoma cells. Such techniques are well known in the art, (e.g., the hybridoma technique originally developed by Kohler and Milstein (Nature 256, 495-497 (1975)) as well as other techniques such as the human B-cell hybridoma technique (Kozbor et al., Immunol. Today 4, 72 (1983)), the EBV-hybridoma technique to produce human monoclonal antibodies (Cole et al., Monoclonal Antibodies in Cancer Therapy (1985) Allen R. Bliss, Inc.,

pages 77-96), and screening of combinatorial antibody libraries (Huse et al., Science 246, 1275 (1989)). Hybridoma cells can be screened immunochemically for production of antibodies specifically reactive with the peptide and the monoclonal antibodies can be isolated. Therefore, the invention also contemplates hybridoma cells secreting monoclonal antibodies with specificity for a glucuronyl transferase of the invention.

The term "antibody" as used herein is intended to include fragments thereof which also specifically react with a protein, of the invention, or peptide thereof. Antibodies can be fragmented using conventional techniques and the fragments screened for utility in the same manner as described above. For example, $F(ab')_2$ fragments can be generated by treating antibody with pepsin. The resulting $F(ab')_2$ fragment can be treated to reduce disulfide bridges to produce Fab' fragments.

Chimeric antibody derivatives, i.e., antibody molecules that combine a non-porcine variable region and a porcine constant region are also contemplated within the scope of the invention. Chimeric antibody molecules can include, for example, the antigen binding domain from an antibody of a mouse, rat, or other species, with porcine constant regions. Conventional methods may be used to make chimeric antibodies containing the immunoglobulin variable region which recognizes a protein of the invention (See, for example, Morrison et al., Proc. Natl Acad. Sci. U.S.A. 81,6851 (1985); Takeda et al., Nature 314, 452 (1985), Cabilly et al., U.S. Patent No. 4,816,567; Boss et al., U.S. Patent No. 4,816,397; Tanaguchi et al., European Patent Publication EP171496; European Patent Publication 0173494, United Kingdom patent GB 2177096B).

Substances that inhibit the expression of the glucuronyl transferase gene include antisense nucleic acid sequences that are complimentary to a sequence of the glucuronyl transferase gene. The antisense sequences of the present invention may be ribonucleic or deoxyribonucleic acids and may contain naturally occurring bases

including adenine, guanine, cytosine, thymidine and uracil. The antisense sequences may also contain modified bases such as xanthine, hypoxanthine, 2-aminoadenine, 6-methyl, 2-propyl and other alkyl adenines, 5-halo uracil, 5-halo cytosine, 6-aza uracil, 6-aza cytosine and 6-aza thymine, pseudo uracil, 4-thiouracil, 8-halo adenine, 8-aminoadenine, 8-thiol adenine, 8-thiolalkyl adenines, 8-hydroxyl adenine and other 8-substituted adenines, 8-halo guanines, 8-amino guanine, 8-thiol guanine, 8-thiolalkyl guanines, 8-hydroxyl guanine and other 8-substituted guanines, other aza and deaza uracils, thymidines, cytosines, adenines, or guanines, 5-trifluoromethyl uracil and 5-trifluoro cytosine. In addition, the antisense sequences of the invention may contain modified phosphorous, oxygen heteroatoms in the phosphate backbone, short chain alkyl or cycloalkyl intersugar linkages or short chain heteroatomic or heterocyclic intersugar linkages. For example, the antisense oligonucleotides may contain phosphorothioates, phosphotriesters, methyl phosphonates, and phosphorodithioates. In an embodiment of the invention there are phosphorothioate bonds links between the four to six 3'-terminus bases. In another embodiment phosphorothioate bonds link all the nucleotides.

The antisense sequences of the invention may also comprise nucleotide analogs that may be better suited as therapeutic or experimental reagents. An example of an oligonucleotide analogue is a peptide nucleic acid (PNA) wherein the deoxyribose (or ribose) phosphate backbone in the DNA (or RNA), is replaced with a polyamide backbone which is similar to that found in peptides (P.E. Nielsen, et al Science 1991, 254, 1497).

2. <u>Screening methods</u>

As hereinbefore mentioned, the present invention provides a method of screening for a substance that regulates skatole metabolism in a pig. Substances that regulate skatole metabolism include substances that modulate the Phase I oxidation or Phase II conjugation reactions involved in skatole metabolism. Preferably, the substances enhance the activity or expression of CYP2E1 or a

sulfotransferase or inhibit the activity or expression of a glucoronyl transferase and are useful in reducing boar taint.

Substances Which Modulate Enzyme Activity

In one aspect, the present invention provides a method of screening for a substance that enhances the activity of CYP2E1 or a sulfotransferase and/or inhibits the activity of a glucuronyl transferase.

(a) CYP2E1

In one embodiment of the invention, a method is provided for screening for a substance that enhances skatole metabolism in a pig by enhancing CYP2E1 activity comprising the steps of:

(a) reacting a substrate of CYP2E1 and CYP2E1, in the presence of a test substance, under conditions such that CYP2E1 is capable of converting the substrate into a reaction product;

(b) assaying for reaction product, unreacted substrate or unreacted CYP2E1;

(c) comparing to controls to determine if the test substance selectively enhances CYP2E1 activity and thereby is capable of enhancing skatole metabolism in a pig.

Substrates of CYP2E1 which may be used in the method of the invention for example include skatole and analogs and derivatives thereof. The corresponding reaction products for skatole are pro-MII and F-1.

The induction of CYP2E1 activity can be measured using a variety of techniques including measuring the levels of the CYP2E1 protein or mRNA or by testing for CYP2E1 activity. CYP2E1 activity can be measured using various assays including assaying for N-nitrosodimethylamine (NDMAD) demethylase activity, aniline hydroxylase activity and p-nitrophenol hydroxylase activity (Xu et al., 1994).

(b) Sulfotransferase

In another embodiment of the invention, a method is provided for screening for a substance that enhances skatole metabolism in a pig by enhancing sulfotransferase activity comprising the steps of:

(a) reacting a substrate of sulfotransferase and sulfotransferase, in the presence of a test substance, under conditions such that sulfotransferase is capable of converting the substrate into a reaction product;

(b) assaying for reaction product, unreacted substrate or unreacted sulfotransferase;

(c) comparing to controls to determine if the test substance selectively enhances sulfotransferase activity and thereby is capable of enhancing skatole metabolism in a pig.

Substrates of sulfotransferase which may be used in the method of the invention for example include pro-MII and analogs and derivatives thereof, and 2-naphthol. The corresponding reaction product for pro-MII is MII.

The induction of sulfotransferase activity can be measured using a variety of techniques including measuring the levels of the sulfotransferase protein or mRNA or by testing for sulfotransferase activity.

(c) Glucuronyl Transferase

In a further embodiment of the invention, a method is provided for screening for a substance that enhances skatole metabolism in a pig by inhibiting glucuronyl transferase activity comprising the steps of:

(a) reacting a substrate of glucuronyl transferase and glucuronyl transferase, in the presence of a test substance, under conditions such that glucuronyl transferase is capable of converting the substrate into a reaction product;

(b) assaying for reaction product, unreacted substrate or unreacted glucuronyl transferase;

(c) comparing to controls to determine if the test substance selectively inhibits glucuronyl transferase activity and thereby is capable of inhibiting skatole metabolism in a pig.

Substrates of glucuronyl transferase which may be used in the method of the invention for example include pro-MII and analogs

and derivatives thereof, 2-naphthol and p-nitrophenol. The corresponding reaction product for pro-MII is MII.

The inhibition of glucuronyl transferase activity can be measured using a variety of techniques including measuring the levels of the glucuronyl transferase protein or mRNA or by testing for glucuronyl transferase activity.

The CYP2E1, sulfotransferase and glucuronyl transferase enzymes used in the method of the invention may be obtained from natural, recombinant, or commercial sources. Cells or liver microsomes expressing the enzymes may also be used in the method.

Conditions which permit the formation of a reaction product may be selected having regard to factors such as the nature and amounts of the test substance and the substrate.

The reaction product, unreacted substrate, or unreacted enzyme; may be isolated by conventional isolation techniques, for example, salting out, chromatography, electrophoresis, gel filtration, fractionation, absorption, polyacrylamide gel electrophoresis, agglutination, or combinations thereof.

To facilitate the assay of the reaction product, unreacted substrate, or unreacted enzyme; antibody against the reaction product or the substance, or a labelled enzyme or substrate, or a labelled substance may be utilized. Antibodies, enzyme, substrate, or the substance may be labelled with a detectable marker such as a radioactive label, antigens that are recognized by a specific labelled antibody, fluorescent compounds, enzymes, antibodies specific for a labelled antigen, and chemiluminescent compounds.

The substrate used in the method of the invention may be insolubilized. For example, it may be bound to a suitable carrier. Examples of suitable carriers are agarose, cellulose, dextran, Sephadex, Sepharose, carboxymethyl cellulose polystyrene, filter paper, ion-exchange resin, plastic film, plastic tube, glass beads, polyamine-methyl vinyl-ether-maleic acid copolymer, amino acid copolymer, ethylene-maleic acid copolymer, nylon, silk, etc. The carrier

may be in the shape of, for example, a tube, test plate, beads, disc, sphere etc. The insolubilized enzyme, substrate, or substance may be prepared by reacting the material with a suitable insoluble carrier using known chemical or physical methods, for example, cyanogen bromide coupling.

Substances which Modulate Gene Expression

In another aspect, the present invention includes a method for screening for a substance that enhances skatole metabolism by modulating the transcription or translation of an enzyme involved in skatole metabolism.

(a) CYP2E1

In one embodiment of the invention, a method is provided for screening for a substance that enhances skatole metabolism by enhancing transcription and/or translation of the gene encoding CYP2E1 comprising the steps of:

(a) culturing a host cell comprising a nucleic acid molecule containing a nuclei acid sequence encoding CYP2E1 and the necessary elements for the transcription or translation of the nucleic acid sequence, and optionally a reporter gene, in the presence of a test substance; and

(b) comparing the level of expression of CYP2E1, or the expression of the protein encoded by the reporter gene with a control cell transfected with a nucleic acid molecule in the absence of the test substance.

(b) Sulfotransferase

In another embodiment of the invention, a method is provided for screening for a substance that enhances skatole metabolism by enhancing transcription and/or translation of the gene encoding sulfotransferase comprising the steps of:

(a) culturing a host cell comprising a nucleic acid molecule containing a nuclei acid sequence encoding sulfotransferase and the necessary elements for the transcription or translation of the nucleic acid sequence, and optionally a reporter gene, in the presence of a test substance; and

(b) comparing the level of expression of sulfotransferase, or the expression of the protein encoded by the reporter gene with a control cell transfected with a nucleic acid molecule in the absence of the test substance.

(c) Glucuronyl Transferase

In a further embodiment of the invention, a method is provided for screening for a substance that enhances skatole metabolism by inhibiting transcription and/or translation of the gene encoding glucuronyl transferase comprising the steps of:

(a) culturing a host cell comprising a nucleic acid molecule containing a nuclei acid sequence encoding glucuronyl transferase and the necessary elements for the transcription or translation of the nucleic acid sequence, and optionally a reporter gene, in the presence of a test substance; and

(b) comparing the level of expression of glucuronyl transferase, or the expression of the protein encoded by the reporter gene with a control cell transfected with a nucleic acid molecule in the absence of the test substance.

A host cell for use in the method of the invention may be prepared by transfecting a suitable host with a nucleic acid molecule comprising a nucleic acid sequence encoding the appropriate enzyme. Suitable transcription and translation elements may be derived from a variety of sources, including bacterial, fungal, viral, mammalian, or insect genes. Selection of appropriate transcription and translation elements is dependent on the host cell chosen, and may be readily accomplished by one of ordinary skill in the art. Examples of such elements include: a transcriptional promoter and enhancer or RNA polymerase binding sequence, a ribosomal binding sequence, including a translation initiation signal. Additionally, depending on the host cell chosen and the vector employed, other genetic elements, such as an origin of replication, additional DNA restriction sites, enhancers, and sequences conferring inducibility of transcription may be incorporated into the expression vector. It will also be appreciated that the necessary

transcription and translation elements may be supplied by the native gene of the enzyme and/or its flanking sequences.

Examples of reporter genes are genes encoding a protein such as β-galactosidase, chloramphenicol acetyltransferase, firefly luciferase, or an immunoglobulin or portion thereof such as the Fc portion of an immunoglobulin, preferably IgG. Transcription of the reporter gene is monitored by changes in the concentration of the reporter protein such as β-galactosidase, chloramphenicol acetyltransferase, or firefly luciferase. This makes it possible to visualize and assay for expression of the enzyme and in particular to determine the effect of a substance on expression of enzyme.

Suitable host cells include a wide variety of prokaryotic and eukaryotic host cells, including bacterial, mammalian, yeast or other fungi, viral, plant, or insect cells. Protocols for the transfection of host cells are well known in the art (see, Sambrook et al. Molecular Cloning A Laboratory Manual, 2nd edition, Cold Spring Harbor Laboratory Press, 1989, which is incorporated herein by reference). Host cells which are commercially available may also be used in the method of the invention. For example, the h2A3 and h2B6 cell lines available from Gentest Corporation are suitable for the screening methods of the invention.

3. Compositions

Substances which enhance skatole metabolism described in detail herein or substances identified using the methods of the invention which selectively enhance CYP2E1 or sulfotransferase activity or inhibit glucuronyl transferase activity (including antibodies or antisense sequences) may be incorporated into pharmaceutical compositions. Therefore, the invention provides a pharmaceutical composition for use in reducing boar taint comprising an effective amount of one or more substances which enhance skatole metabolism and/or a pharmaceutically acceptable carrier, diluent, or excipient. In one embodiment, the present invention provides a pharmaceutical

composition comprising an effective amount of the substance which is selected from the group consisting of

(a) a substance that increases the activity of the CYP2E1 enzyme;

(b) a substance that induces or increases the expression of the CYP2E1 gene;

(c) a substance that increases the activity of the sulfotransferase enzyme;

(d) a substance that induces or increases the expression of the sulfotransferase gene;

(e) a substance that decreases the activity of the glucuronyl transferase enzyme; and

(f) a substance that reduces or decreases the expression of the glucuronyl transferase gene.

The substances for the present invention can be administered for oral, topical, rectal, parenteral, local, inhalant or intracerebral use. Preferably, the active substances are administered orally (in the food or drink) or as an injectable formulation.

In the methods of the present invention, the substances described in detail herein and identified using the method of the invention form the active ingredient, and are typically administered in admixture with suitable pharmaceutical diluents, excipients, or carriers suitably selected with respect to the intended form of administration, that is, oral tablets, capsules, elixirs, syrups and the like, consistent with conventional vetrinary practices.

For example, for oral administration the active ingredients may be prepared in the form of a tablet or capsule for inclusion in the food or drink. In such a case, the active substances can be combined with an oral, non-toxic, pharmaceutically acceptable, inert carrier such as lactose, starch, sucrose, glucose, methyl cellulose, magnesium stearate, dicalcium phosphate, calcium sulfate, mannitol, sorbitol and the like; for oral administration in liquid form, the oral active substances can be combined with any oral, non-toxic,

pharmaceutically acceptable inert carrier such as ethanol, glycerol, water, and the like. Suitable binders, lubricants, disintegrating agents, and coloring agents can also be incorporated into the dosage form if desired or necessary. Suitable binders include starch, gelatin, natural sugars such as glucose or beta-lactose, corn sweeteners, natural and synthetic gums such as acacia, tragacanth, or sodium alginate, carboxymethylcellulose, polyethylene glycol, waxes, and the like. Suitable lubricants used in these dosage forms include sodium oleate, sodium stearate, magnesium stearate, sodium benzoate, sodium acetate, sodium chloride, and the like. Examples of disintegrators include starch, methyl cellulose, agar, bentonite, xanthan gum, and the like.

Gelatin capsules may contain the active substance and powdered carriers, such as lactose, starch, cellulose derivatives, magnesium stearate, stearic acid, and the like. Similar carriers and diluents may be used to make compressed tablets. Tablets and capsules can be manufactured as sustained release products to provide for continuous release of active ingredients over a period of time. Compressed tablets can be sugar coated or film coated to mask any unpleasant taste and protect the tablet from the atmosphere, or enteric coated for selective disintegration in the gastrointestinal tract. Liquid dosage forms for oral administration may contain coloring and flavoring agents to increase acceptance.

Water, a suitable oil, saline, aqueous dextrose, and related sugar solutions and glycols such as propylene glycol or polyethylene glycols, may be used as carriers for parenteral solutions. Such solutions also preferably contain a water soluble salt of the active ingredient, suitable stabilizing agents, and if necessary, buffer substances. Suitable stabilizing agents include antioxidizing agents such as sodium bisulfate, sodium sulfite, or ascorbic acid, either alone or combined, citric acid and its salts and sodium EDTA. Parenteral solutions may also contain preservatives, such as benzalkonium chloride, methyl- or propyl-paraben, and chlorobutanol.

The substances described in detail herein and identified using the methods of the invention can also be administered in the form of liposome delivery systems, such as small unilamellar vesicles, large unilamellar vesicles, and multilamellar vesicles. Liposomes can be formed from a variety of phospholipids, such as cholesterol, stearylamine, or phosphatidylcholines.

Substances described in detail herein and identified using the methods of the invention may also be coupled with soluble polymers which are targetable drug carriers. Examples of such polymers include polyvinylpyrrolidone, pyran copolymer, polyhydroxypropyl-methacrylamidephenol, polyhydroxyethyl-aspartamidephenol, or polyethyleneoxide-polylysine substituted with palmitoyl residues. The substances may also be coupled to biodegradable polymers useful in achieving controlled release of a drug. Suitable polymers include polylactic acid, polyglycolic acid, copolymers of polylactic and polyglycolic acid, polyepsilon caprolactone, polyhydroxy butyric acid, polyorthoesters, polyacetals, polydihydropyrans, polycyanoacylates, and crosslinked or amphipathic block copolymers of hydrogels.

Suitable pharmaceutical carriers and methods of preparing pharmaceutical dosage forms are described in Remington's Pharmaceutical Sciences, Mack Publishing Company, a standard reference text in this field.

More than one substance described in detail herein or identified using the methods of the invention may be used to enhance metabolism of skatole. In such cases the substances can be administered by any conventional means available for the use in conjunction with pharmaceuticals, either as individual separate dosage units administered simultaneously or concurrently, or in a physical combination of each component therapeutic agent in a single or combined dosage unit. The active agents can be administered alone, but are generally administered with a pharmaceutical carrier selected on the basis of the chosen route of administration and standard pharmaceutical practice as described herein.

4. Genetic Screening

The present invention further includes the identification of polymorphisms in genes encoding the enzymes responsible for skatole metabolism in a pig including CYP2E1, sulfotransferases and glucuronyl transferase as described in detail hereinabove. The identification of genes that encode these enzymes from pigs that are high skatole metabolizers (and hence have a low incidence of low boar taint) can be used to develop lines of pigs that have a low incidence of boar taint. In addition, the identification of these genes can be used as markers for identifying pigs that are predisposed to having a low incidence of boar taint.

Accordingly, the present invention provides a method for producing pigs which have a lower incidence of boar taint comprising selecting pigs that express high levels of CYP2E1 and/or sulfotransferase and/or low levels of glucuronyl transferase; and breeding the selected pigs.

Transgenic pigs may also be prepared which produce high levels of CYP2E1 and/or sulfotransferase and/or low levels of glucuronyl transferase. The transgenic pigs may be prepared using conventional techniques. For example, a recombinant molecule may be used to introduce (a) a gene encoding CYP2E1 or (b) a gene encoding a sulfotransferase or (c) an antisense nucleic acid molecule complimentary to a glucuronyl transferase. Such recombinant constructs may be introduced into cells such as embryonic stem cells, by a technique such as transfection, electroporation, injection, etc. Cells which show high levels of CYP2E1 and/or sulfotransferase and/or low levels of glucuronyl transferase may be identified for example by Southern Blotting, Northern Blotting, or by other methods known in the art. Such cells may then be fused to embryonic stem cells to generate transgenic animals. Germline transmission of the mutation may be achieved by, for example, aggregating the embryonic stem cells with early stage embryos, such as 8 cell embryos, transferring the resulting blastocysts into recipient females in vitro, and generating germline

transmission of the resulting aggregation chimeras. Such a transgenic pig may be mated with pigs having a similar phenotype i.e. producing high levels of CYP2E1 and/or sulfotransferase and/or low levels of glucuronyl transferase to produce animals having a low incidence of boar taint.

The following non-limiting examples are illustrative of the present invention:

EXAMPLES

Example 1

Cytochrome P450IIE1 Oxidation Reactions in Skatole Metabolism

Materials and Methods

Chemicals

Skatole (3-methylindole), indole-3-acetonitrile, NADH (disodium salt), NADPH (tetrasodium salt), diallyl sulfide (DAS), chlorzoxazone and metyrapone were purchased from Sigma Chemical (St. Louis, MO). SKF 525A hydrochloride (Proadifen) was purchased from Research Biochemical Inc. (Natick, MA). 6-Hydroxy-3-methylindole (pro-MII) and 3-hydroxy-3-methyloxindole (MIII) were kindly provided by Jens Hansen Møller of the Danish Meat Research Institute, Roskilde, Denmark.

Animals

Intact male pigs obtained by back-crossing F3 European Wild Pig x Swedish Yorkshire pigs with Swedish Yorkshire sows (Anderson et al., 1994; Squires and Lundström, 1997) were used. They were slaughtered at approximately 108 kg live weight. Backfat samples were collected and analyzed for skatole concentration with a colorimetric assay (Mortensen and Sørensen, 1984). Liver samples were frozen in liquid nitrogen for subsequent preparation of microsomes and analysis of levels of P450IIE1 (Squires and Lundström, 1997) and for analysis of skatole metabolism. On the basis of a preliminary analysis of skatole metabolism by liver microsomes, 9 pigs with sufficient level of production of skatole metabolites for HPLC analysis were chosen. Concentrations of P450IIE1 in the liver of these pigs were medium to

high (mean, 62.1; SD, 32.4 of the arbitrary units used by Squires and Lundström, 1997), while concentrations of skatole in the backfat were low (mean, .14 ; SD, .04 ppm).

Preparation of Microsomes

Liver microsomes were prepared as previously described by Meadus and Squires (1995).

Enzyme Assays

The metabolism of skatole was assayed in a medium containing 50 mM Tris-HCL buffer, pH 7.4, 10 mM potassium phosphate, .1 mM EDTA, 20% glycerol, .01 mM skatole and 1 mg/mL microsomal protein in a total volume of 2 mL. The reactions were started by addition of NADH and NADPH to 1mM final concentration of each after 3 min of preincubation at 37°C. After 30 min at 37°C, the reactions were terminated by addition of 2 mL of ice-cold .2 M ammonium acetate buffer, pH 5.0.

Those incubations with no inhibitor added were regarded as controls. The effect of different P450 inhibitors on skatole metabolism was evaluated under the same conditions by including different concentrations of P450 inhibitors into the medium before preincubation. The concentrations of the inhibitors ranged from .025 to .2 mM of SKF 525A, diallyl sulfide (DAS) or chlorzoxazone, or .025 to .1 mM of metyrapone. At higher inhibitor concentrations up to .4 mM, no significant changes in the formation of metabolites or skatole disappearance were observed.

Each assay was performed in duplicate. The assays of formation of skatole metabolites over time were performed with microsomes from 2 different randomly chosen pigs. The control incubations and the incubations with P450 inhibitors were performed with microsomes from 3 different randomly chosen pigs.

Analysis of skatole metabolites

The procedures for extraction and analysis of skatole metabolites by HPLC were based on the method described by Bæk et al. (1995). After adding ammonium acetate buffer, 5 µL of .01 mg/mL

indole-3-acetonitrile was added to the reaction mixture as an internal standard. The mixture was then passed through a solid phase extraction tC18 cartridge (Waters Co., Milford, MA) previously conditioned with 2 mL of methanol and then 2 mL of water. The column was washed with 1 mL of water and skatole metabolites were eluted with 2 x .5 mL of acetonitrile. The acetonitrile was removed under a gentle stream of nitrogen at 40°C and the metabolites were resuspended in 200 µl of acetonitrile for HPLC analysis. The HPLC equipment included a Spectra-Physics (San Jose, CA, USA) SP8800 pump, SP8800 autosampler and SP4290 integrator, and a Phenomenex (Torrance, CA, USA) Prodigy ODS-2 C18, 250 x 4.6 mm , 5 µm column. The analysis was performed using 3 buffers, A - acetonitrile: .01 mM potassium phosphate buffer, pH 3.9 (10:90), B - acetonitrile:water (90:10) and C - acetonitrile, with the following gradient: 0 min - A, 100%; 6 min - A, 75%, B, 25%; 15min - A, 75%, B, 25%; 18 min - A, 20%, B, 80%; 20 min - B, 100%; 23 min - C, 100%; 25 min - C, 100%; 26 min - A, 100%; 30 min A, 100%. Injection volume was 100 µl. Skatole metabolites were detected simultaneously using both a UV detector set at 250 nm (Spectra 100, Spectra-Physics, San Jose, CA, USA) and fluorescence detector (Shimadzu RF-535; Shimadzu Co., Kyoto, Japan), excitation wavelength of 285 nm and emission wavelength of 340 nm.

Statistical analysis

The effect of different concentrations of P450 inhibitors on the concentration of skatole and its metabolites in microsomal incubations was analyzed with Statistical Analysis System (SAS Institute, 1995) using GLM procedure. For the purpose of the analysis, peak areas of the metabolites in control incubations were considered as 100%. Peak areas of the metabolites in the incubations with inhibitors were transformed to % values of those for controls. The statistical model included the effect of the concentration of inhibitor and the effect of animal.

Results

A HPLC profile of microsomal metabolites of skatole is shown in Figure 1. Several microsomal metabolites of skatole were found; the peaks F-4 and UV-2 were identified as 6-hydroxyskatole (pro-MII), which is a precursor of its sulfated form, 6-sulfatoxyskatole (MII), and 3-hydroxy-3-methyloxindole (MIII), respectively, using standards. Other metabolites were not identified in this study. The production of skatole metabolites by liver microsomes was approximately linear for 30 min (Figure 2) and this period of time was chosen for subsequent assays using cytochrome P450 inhibitors.

The microsomal assays which included addition of SKF 525A (Figure 3) or metyrapone (Figure 4) indicated a significant effect ($P = .001$) on pro-MII synthesis. The synthesis of pro-MII was decreased with .025 mM of added inhibitor ($P < .05$; Table 1), decreasing to 38.2% with .2 mM of SKF 525A and 11.6% with .1 mM of metyrapone compared to controls. Formation of MIII was inhibited by SKF 525A ($P < .001$). The effect of metyrapone on MIII could not be evaluated, because a metabolite of metyrapone co-eluted with MIII in the HPLC analysis. Both inhibitors reduced formation of UV-3. SKF 525A also reduced formation of F-2 and F-3 ($P < .05$). The amount of skatole remaining in the reaction mixture after incubation was higher for the assays with metyrapone than for control ($P = .008$), and not affected by SKF 525A ($P = .613$).

The specific inhibitors of P450IIE1 were more effective in reducing skatole metabolism and formation of its metabolites than was SKF 525A and metyrapone. The synthesis of pro-MII was significantly reduced with .025 mM of DAS or chlorzoxazone, while at .2 mM of these inhibitors the synthesis of MII was reduced to 30.9% and 9.7%, respectively. Both inhibitors reduced MIII synthesis and skatole disappearance (Table 1). DAS also inhibited the formation of all the other skatole metabolites ($P = .001$), except for F-1 ($P = .336$; Figure 5). Chlorzoxazone inhibited the formation of all skatole metabolites ($P < .05$), except for F-3 ($P = .680$; Figure 6).

Discussion

In this study, the involvement of the liver enzyme system, cytochrome P450, in the biodegradation of skatole to pro-MII and other metabolites was evaluated.

In this study, microsomal incubations were conducted using standard procedures for evaluating P450 activity, using NADPH and NADH as electron donors. These incubations revealed formation of several skatole metabolites, including pro-MII and MIII, detected by either fluorescence or UV absorbance after separation on HPLC (Figure 1 and 2). Besides pigs, MIII has been also found in mouse urine (Skiles et al., 1989) and, in a conjugated form, in goat urine (Smith et al., 1993). Pro-MII or MII has not yet been reported in other species than pigs.

The formation of MII in microsomal incubations was inhibited by both classical inhibitors of cytochrome P450, metyrapone and SKF 252A. These inhibitors form ligand complexes with cytochrome P450, causing spectral changes of the enzymes (Netter 1980). Metyrapone forms complexes with variety of P450 isozymes including P4502E1 (Knecht et al., 1993), by binding competitively to P450 iron binding site II preventing oxidation of a substrate (Netter, 1980). However, the affinity of metyrapone towards different forms of P450 is substantially different (Roos et al., 1993) and some of the isozymes are not affected by this compound (Ryan et al., 1984). These could be the reasons for a strong inhibitory effect of metyrapone on pro-MII formation, reduction of UV-3 formation and skatole disappearance, but no effect on the formation of the other metabolites. SKF 252A is considered a potent inhibitor of cytochrome P450, acting both as a competitive inhibitor binding to cytochrome P450 at its lipophilic site I, and as a non-competitive inhibitor (Netter, 1980). Although a different potency for some forms of P450 has been reported, SKF 525A is considered a strong inhibitor of the whole cytochrome P450 system (Murray and Reidy, 1990). In this study, SKF 525A inhibited the formation of pro-MII, MIII and 3 other skatole metabolites. These results show the involvement of cytochrome P450 in oxidative metabolism of

skatole in pig liver. Particularly, the metabolic pathway leading to formation of MII was strongly affected by both general cytochrome P450 inhibitors.

The next step of this study was to investigate the role of a P450 isozyme, P450IIE1, in skatole metabolism. This enzyme is involved in both oxidative as well as reductive metabolism. A number of substrates of P450IIE1 has been described (Koop, 1992). They include both polar and non-polar compounds of different molecular structures but with a small size. This led to a suggestion that the active site of P450IIE1 acts as a molecular sieve (Guengerich and Shimada, 1991). The involvement of P450IIE1 in skatole metabolism in pigs has been suggested by a recent report of a relationship between the ability of pig liver microsomes to metabolize skatole to MII and the ability to oxidize chlorzoxazone, a specific substrate for P450IIE1 (Friis, 1995). Also, a negative correlation (r = -.68) between fat skatole concentrations and liver concentrations of P450IIE1 determined by Western blotting pointed this enzyme as being responsible for skatole metabolism (Squires and Lundström, 1997). Chlorzoxazone and DAS were used as inhibitors specific for cytochrome P450IIE1. Chlorzoxazone is a competitive inhibitor of P450IIE1 (Peter et al., 1990). Chlorzoxazone hydroxylation is also catalyzed by P450 1A1 and 3A, but only at small substrate concentrations (2 μM; Jayyosi et al., 1995). At higher concentrations (.2 mM), chlorzoxazone is metabolized almost exclusively by P450IIE1. DAS is both a competitive and via its metabolite, DASO2, a suicidal inhibitor of P450IIE1 (Brady et al., 1991). DAS has not been reported to inhibit other P450 isozymes.

Both chlorzoxazone and DAS inhibited skatole disappearance and formation of nearly all skatole metabolites in microsomal incubations, indicating P450IIE is involved in the synthesis of these metabolites. This confirms the earlier indications that skatole metabolism in pig liver is catalyzed by cytochrome P450IIE1. Whether or not other forms of cytochrome P450 are also involved in skatole metabolism can not be concluded from this study. Thornton-Manning

et al. (1996) reported that eight other forms of P450 besides P450IIE1 can metabolize skatole. However, they used vaccinia-expressed human and rodent P450s and the synthesis of pro-MII was not evaluated. The strong inhibitory effect of specific P450IIE1 inhibitors compared to general P450 inhibitors on the formation of most skatole metabolites found in this study suggests that cytochrome P450IIE1 is a major enzyme responsible for skatole metabolism in pigs. Moreover, the strong effect of both DAS and chlorzoxazone on pro-MII synthesis indicates that this enzyme is involved in the formation of this important metabolite of skatole.

Skatole is absorbed from the gut and metabolized in the liver and skatole which remains in the blood is quickly deposited in the fatty tissue. It is also relatively rapidly released from the fat into the blood stream and excreted in the urine in the metabolized form (Friis, 1993). Thus, the activity of P450IIE1 in the liver, which converts skatole into metabolites for excretion, is likely to have a dramatic effect on fat skatole concentrations in pigs. It is particularly so because cytochrome P450IIE1 is involved in the production of MII.

The present results strongly suggest that cytochrome P450IIE1 catalyzes the biotransformation of skatole in pig liver to pro-MII, an important skatole metabolite, as well as to other metabolites. The metabolism of skatole in the liver by this enzyme could be an important factor in regulating the concentrations of skatole in the fat of pigs. An understanding of this mechanism is important for controlling boar taint and consequently enabling the use of intact males for pork production.

Example 2

Conjugation Reactions in Skatole Metabolism

Materials and Methods

Chemicals

Skatole (3-methylindole), indole-3-acetonitrile, 2-naphthol, p-nitrophenol, PAPS (adenosine 3'-phosphate 5'-phosphosulfate), UDPGA (uridine 5'-diphosphoglucuronyl, trisodium salt), NADPH and NADH were purchased from Sigma Chemical Co. (St.

Louis, MO, USA). 6-Hydroxy-3-methylindole (pro-MII) and 3-hydroxy-3-methyloxindole (MIII) were graciously provided by Jens Hansen Møller of the Danish Meat Research Institute, Roskilde, Denmark.

Animals

Trial I. Trial I included Yorkshire intact male pigs from a line selected for reduced backfat thickness and increased growth rate (McKay, 1990). A total of 18 pigs were selected based on the levels of skatole in backfat measured by a colorimetric procedure (Mortensen and Sørensen, 1984) . The low group included 9 males with mean skatole levels of .06, SD, .02, and range of .02 to .08 ppm. The high group consisted of 9 males with mean fat skatole levels of .42, SD, .26 and range of .27 to 1.07 ppm. The pigs were fed a barley, wheat and soybean meal diet containing 16% CP, 13.4 MJ DE/kg and 4% crude fiber and were slaughtered at approximately 100 kg live weight.

Trial II. Trail II included a total of 45 F4 European Wild Pig x Swedish Yorkshire intact male pigs developed as described elsewhere (Squires and Lundström, 1997) were used. They were fed barley, oat and soybean meal diet containing 16.8% CP, 12.3 MJ DE/kg and 4.4% crude fiber and were slaughtered following stunning with carbon dioxide at approximately 108 kg live weight. The levels of skatole in backfat were measured by a colorimetric procedure (Mortensen and Sørensen, 1984). Liver samples, taken at slaughter, were frozen in liquid nitrogen and subsequently analyzed for levels of P450IIE1 by Western blotting (Squires and Lundström, 1997). Liver samples from 22 males selected for a wide range of the levels of P450IIE1 in the liver and skatole in the fat were used for enzymatic assays.

Throughout the experiments the pigs were treated according to accepted standards for humane treatment of animals.

Preparation of Microsomes and Homogenates

Liver microsomes were prepared as described by Meadus and Squires (1995). Tissues were prepared for sulfation assays in Trial 1 by homogenizing liver samples in 5 volumes of 50 mM Tris-HCL buffer,

pH 7.4, 100 mM KCL, 10 mM EDTA followed by centrifugation at 10,000 x g for 20 min. The supernatant was used for further study. Protein content in both microsomes and homogenates were measured with BCA Protein Assay (Pierce, Rockford, IL, USA).

Enzyme Assays

Trial I. The rate of skatole metabolism by liver microsomes was assayed in a medium containing 50 mM Tris-HCL buffer, pH 7.4, 10 mM potassium phosphate, .1 mM EDTA, 20% glycerol (Buffer I), with .01 mM skatole and 1mg/mL microsomal protein in a total volume of 100:l. The incubations were started, after 3 min preincubation at 37°C, by addition of 1mM NADPH and 1mM NADH. The reaction was terminated after 10 min by addition of 100 µl of ice cold methanol. The mixture was vortexed and centrifuged for 5 min at 2000 x g to precipitate protein. Supernatant (100 µl) was analyzed by HPLC and the rate of skatole metabolism was evaluated by monitoring the changes in skatole concentration.

The activities of UDP-glucuronyltransferase were assayed in 50 mM Tris-HCL buffer, pH 7.4, 10 mM MgCl2, .1 mM EDTA (Buffer II). The assays with p-nitrophenol contained .06 mM p-nitrophenol and 1mg/mL microsomal protein, in a total volume of 1 mL; the assays with 2-naphthol contained .02 mM 2-naphthol and .1 mg/mL microsomal protein in a total volume of 100:l. Both assays were started by addition of .5 mM UDPGA and were performed at 37°C for 5 min. The activity of sulfotransferase was assayed using 2-naphthol as a substrate. The incubation mixture contained Buffer II, from 1.9 to 3.1 mg/ml protein, .01 mM 2-naphthol and 1 mM PAPS, in a total volume of 100 :l. These incubations were performed at 37°C for 60 min. The concentration of p-nitrophenol was determined by measuring the absorbance of the reaction mixture at 400n immediately after incubation. The assays with 2-naphthol were terminated by addition of 100:l of ice-cold methanol, centrifuged and the concentration of 2-naphthol in the supernatant was measured by HPLC as described for the assays of skatole metabolism in microsomes.

Trial II. For evaluation of skatole metabolite production (Trial II-a, n = 22), 1mg/ml of liver microsomes were incubated in Buffer I, 1mM NADPH, 1mM NADH, and .01 mM skatole in a total volume of 2 ml for 30 min at 37°C as previously described (Babol et al., 1997). To determine which microsomal skatole metabolites are conjugated with sulfate (Trial II-b, n = 3), the reaction mixture was first incubated for 30 min as in Trial II-a. PAPS was then added to 1 mM final concentration and the incubation continued for 30, 60 and 90 min. Control incubations were performed for the same period of time with no PAPS added. Assays to evaluate the conjugation of skatole metabolites with glucuronyl (Trial II-c, n = 3) were conducted in a similar way by adding 1 mM of UDP-GA after 30 min of initial incubation and continuing the incubation for a further 7.5, 15 and 30 min. For conjugation assays in Trial II-b and II-c liver microsomes from pigs with sufficient levels of production of skatole metabolites were used. The mean levels of skatole in the fat and cytochrome P450IIE1 in those samples were, in Trial II-b, .11 (SD, .03 and 62.4 (SD, 41.9), respectively, and, in Trial II-c, 10 (SD, .04) and 101.1 (SD, 30.2), respectively.

The rate of pro-MII glucuronidation (Trial II-d, n = 22) was assayed in Buffer II containing .1mg/ml microsomal protein, .5 mM UDPGA and approximately .5 µg/ml of pro-MII in a total volume of 2 ml. The assays were incubated at 37°C for 5 min. The rate of pro-MII sulfation by liver microsomes (Trial II-e, n = 22) was evaluated in Buffer II containing 3 mg/ml of liver microsomes, 1mM PAPS and approximately 1 µg/ml pro-MII in a total volume of .5 ml. These incubations were conducted at 37°C for 30 min. The rate of conjugation was calculated from the amount of the products of the conjugation reaction that were formed. It was observed that pro-MII was rapidly metabolized in microsomal incubations when no PAPS was added and the rate of this metabolism varied among the samples. Therefore, parallel incubations were performed with no PAPS added (Trial II-f, n = 22). The rate of pro-MII disappearance in these incubation was defined as pro-MII degradation. Since pro-MII is somewhat unstable, the HPLC

analysis was performed the same day that the incubations were carried out.

All assays in Trial II were terminated by adding an equal amount of .2 M ammonium acetate buffer, pH 5.0, and 5 µl of .01mg/ml indole-3-acetonitrile was added as an internal standard. The metabolites were concentrated by solid phase extraction for HPLC analysis as described elsewhere (Babol et al., 1997). All assays were performed in duplicate.

HPLC

Trial I. The analysis of skatole and 2-naphthol concentrations was performed by a modified HPLC method of Hansen-Møller (1992).The modifications included a different gradient profile: 0 min - buffer A, 100%; 3min - A, 85%; 11 min - A85%; 13 min - A, 20%; 15 min - A, 0%; 18 min - A, 0%; 18.5 min - A, 100%; 23 min - A, 100% and an addition of a guard column. The HPLC equipment included Spectra-Physics (San Jose, CA, USA) SP8800 pump and autosampler, and SP4290 integrator, a Supelcosil RPLC-18, 25 cm x 4.6 I.D. particle size 5 :m column (Supelco Canada Inc., Oakville, ON, Canada) and packed with the same material 4 cm x 4.6 I.D. guard column. Both skatole and 2-naphthol were detected using Varian 2070 spectrofluorometer (Varian Canada Inc,. London, ON, Canada) with excitation wavelength set at 285 nm and emission wavelength set at 340 nm.

Trial II. HPLC procedures used in Trial II are described in detail elsewhere (Babol et al., 1997). The metabolites were separated with RP C18 column and detected by measuring UV absorbance at 250 nm and fluorescence with excitation and emission wavelengths set at 285 and 340 nm.

Statistical analysis

All data were analyzed with the Statistical Analysis System (SAS Institute, 1995). The concentrations of skatole metabolites are expressed in arbitrary units of peak area derived from HPLC analysis. For the purpose of the analysis, some of the measurements were

transformed (see below) to correct for their skewed distribution. However, all values given within the text are from untransformed data. The ratio of glucuronidation:sulfation activity was calculated in both trials to account for the competitive character of both reactions, and the ratio of pro-MII:MIII formation was calculated in Trial II to determine if it is related to fat skatole levels.

Trial I. Skatole levels in fat, skatole metabolism rate and the ratio of 2-naphthol glucuronidation:2-naphthol sulfation were transformed to common logarithms, and 2-naphthol sulfation to square root values. The differences between the pigs with high and low fat skatole levels and the rate of skatole metabolism and conjugation activities were analyzed with ANOVA using the GLM procedure.

Trial II. The effect of time of incubation of skatole metabolites with PAPS (Trial II-b) and UDPGA (Trial II-c) on the concentrations of the metabolites was analyzed using untransformed values with the GLM procedure. The model included the effect of time and sample. For the analysis of the results of Trials II-a, -d, -e and -f the following values were transformed to common logarithms: fat skatole levels, the peak areas of skatole metabolites, pro-MII, F-2, F-3, MIII, UV-1, the ratio of MII:MIII, pro-MII sulfation, the ratio of pro-MII glucuronidation:sulfation and pro-MII degradation. Stepwise regression analysis was used to evaluate the effect of cytochrome P450IIE1 and various skatole metabolites on the levels of skatole in fat. In addition, the pigs were divided according to P450IIE1 levels into 3 groups, low, medium and high. The differences in the metabolic activities among the groups were analyzed with the GLM procedure.

Results

Trial I

The intact male pigs with high fat skatole levels had a higher rate of skatole metabolism, higher activity of UDP-glucuronyltransferase towards 2-naphthol and higher ratio of 2-naphthol glucuronidation: 2-naphthol sulfation ($P < .05$, Table 2) than did males with low levels of skatole in fat. The activity of

UDP-glucuronyltransferase towards p-nitrophenol did not differ between the treatments. The activities of sulfotransferase towards 2-naphthol were decreased in the high as compared to the low fat skatole group. Skatole levels in fat were correlated with the rate of skatole metabolism, r = .57, P = .013 and the rate of glucuronidation of 2-naphthol, r = .52, P = .028. The ratio of 2-naphthol glucuronidation:sulfation was more closely correlated to 2-naphthol sulfation (r = -.87, P = .001) than to glucuronidation (r = .54, P = .02) .

Trial II.

Skatole metabolites evaluated in Trials II-a, -b and -c of this experiment were described elsewhere (Babol et al., 1997). They included pro-MII, MIII, and 5 others not identified skatole microsomal metabolites denoted as F-1, F-2, F-3, UV-1 and UV-2.

The conjugation of microsomal metabolites of skatole over time with glucuronyl (Trial II-c) and sulfate (Trial II-b) are shown in Figures 7 and 8. In Trial II-c, only the concentrations of pro-MII and F-1 were lower than those in control incubations (P < .05). Mean levels of pro-MII and F-1 were both decreased after 7.5 min, and after 30 min reached 20% (SD, 25.2) and 77.8% (SD, 6.9), respectively, of those in control incubations. The mean concentrations of F-2 in 15 and 30 min incubations were numerically low (mean, 68.7%; SD, 60.2 and mean , 44.6%; SD, 39.2, respectively) but not significantly different from control (P = .103). The concentration of skatole was increased in the assays with UDPGA to 126.1% (SD, 39.2) of that for control in 30 min incubation (P = .008). In Trial II-b only the concentration of pro-MII was affected (P = .028) and decreased to 40% (SD, 30.3) of that for control after incubating for 90 min. The mean concentrations of F-2, although numerically several times higher in sulfation assays, did not significantly differ from controls (P = .274).

Incubation of pro-MII with liver microsomes and UDPGA (Trial II-d) resulted in production of two pro-MII metabolites (Figure 9a). Because these were not well separated by HPLC, the sum of the peak areas of both metabolites were used to quantify the

glucuronidation activity towards pro-MII. In Trial II-e, the activity of pro-MII sulfation was determined by quantification of the single product of pro-MII sulfation (Figure 9b). The retention time from HPLC of this metabolite was the same as that of F-2 in Trial II-a, suggesting that this might be the same metabolite. In addition, the formation of this metabolite showed a strong tendency to increase in Trial II-b. The formation of this metabolite tended to be lower than in the control in Trial II-c. If F-2 is a product of pro-MII sulfation the tendency to decrease its formation observed in Trial II-b might be due to a reduced sulfation rather than to glucuronidation of pro-MII.

The results of the ANOVA analysis of the pigs divided into low (L), medium (M) and high (H) groups according to the levels of P450IIE1 in the liver are summarized in Table 3. The L pigs compared to both M and H pigs had a lower ($P < .05$) rate of formation of F-1 and sulfation of pro-MII, and higher ($P < .05$) levels of skatole in fat, rate of UV-1 formation, pro-MII degradation, and the ratio of pro-MII glucuronidation:sulfation.

The levels of skatole in fat were negatively correlated with the levels of cytochrome P450IIE1, metabolite F-1, sulfation of pro-MII, and positively correlated with the ratio of pro-MII glucuronidation:sulfation (Table 4). The correlation between fat skatole and liver P450IIE1 levels ($r = -.79$, $P < .001$) was higher in this study than that reported previously for a larger number of the same pigs ($r = -.68$; Squires and Lundström, 1997) because the pigs used here were selected for a wide variation in P450IIE1 and skatole levels. The levels of P450IIE1 were correlated with the rate of pro-MII sulfation and negatively correlated with the ratio of pro-MII glucuronidation:sulfation but not related to the rate of skatole metabolism or formation of skatole metabolites. Skatole oxidative metabolites were correlated with each other and with pro-MII conjugation products to various degrees. The ratio of pro-MII glucuronidation:sulfation was more closely correlated with pro-MII sulfation ($r = -.97$) than with pro-MII glucuronidation ($r = .67$).

Stepwise regression analysis revealed an effect on fat skatole levels of both P450IIE1 (P = .001) and metabolite F-1 (P = .028). The R2 for this model was .71, compared to .62 for P450IIE1 alone. No other variable was included in the model at the P < .05 rejection level.

Discussion

This study was conducted to investigate relationships among the levels of cytochrome P450IIE1, skatole metabolism, the formation of various skatole metabolites in the liver and the levels of skatole in fat. The rate of skatole metabolism by liver microsomes in pigs in this study was lower than those reported for humans and goats (Ruangyuttckarn et al., 1991). The rate of skatole metabolism was increased in pigs with high fat skatole levels and positively correlated with fat skatole levels in Trial 1, and not related to fat skatole levels in Trial 2. This indicates that the overall rate of skatole metabolism in the liver is not related to skatole clearance and deposition in fat. The positive relation between skatole metabolism rate and fat skatole levels found in Trial 1 could be a result of a substrate stimulation of the P450IIE1 activity as it is the case for a number of other chemicals (Koop, 1992). However, injecting pigs daily for 9 days with high doses of skatole did not increase skatole metabolism rate (Babol and Squires, unpublished data). Among oxidative skatole metabolites only the formation of F-1 was correlated with fat skatole levels (r = -.59), suggesting that the ability to produce this metabolite could be important in the metabolic clearance of skatole. When included with P450IIE1 in the multiple regression model, 71% of variation in fat skatole levels was explained by this model, leaving only 29% to other factors such as diet or environmental conditions, which are known to be important (Kjeldsen, 1993). However, the effect of P450IIE1 may be overestimated in this study since the pigs were selected for a wide variation in the levels of this enzyme. Grouping the pigs according to cytochrome P450IIE1 levels in the liver showed that fat skatole levels were higher in the pigs with low (L) and similar in the pigs with medium (M) and high (H) P450IIE1 levels. F-1 formation was increased in the M and H pigs compared to L

pigs, further confirming its possible role in effective skatole clearance. The higher concentration of UV-1 in liver microsomal incubations from L than from M and H pigs indicate that the formation of this metabolite is related to a slow clearance of skatole, possibly because this metabolite is not easily excreted. Formation of pro-MII, MIII and the ratio of pro-MII:MIII were not correlated to fat skatole levels. The variation in the ratio of pro-MII:MIII almost exclusively depended on the variation in the pro-MII formation, judging from the close correlation of the ratio with MII (r = .94), compared to MIII (r = -. 50). The formation of MIII seems to be irrelevant in the rate of skatole clearance, since it was formed at a similar rate in all L, M and H pigs.

On the other hand, the results of the conjugation assays strongly suggest that the subsequent conjugation of pro-MII with sulfate or glucuronyl are important in the metabolic clearance of skatole in pigs. Pro-MII was the only metabolite of skatole that readily formed conjugates with both sulfate and glucuronyl. F-1 also formed conjugates but only with glucuronyl, while skatole and all the other metabolites did not react with either glucuronyl or sulfate. Sulfation and glucuronidation are major pathways of metabolism of endogenous and exogenous compounds, such as steroids, bile acids, neurotransmitters, drugs and xenobiotics (Tephly and Burchell, 1990; Falany, 1991). Pro-MII and all the other skatole metabolites found in plasma or urine, with the exception of MIII are in the conjugated form (Bæk, 1995), indicating that conjugation reactions are necessary for clearance of skatole. In this study, distinct differences in conjugation activities in the liver were found in pigs with different skatole levels in fat. In Trial I, the activities of sulfotransferase and UDP-glucuronyltransferase were investigated using common substrates. p-Nitrophenol was used for evaluating glucuronidation and 2-naphthol was used for evaluating both glucuronidation and sulfation activities. Both p-nitrophenol and 2-naphthol are phenolic compounds predominantly used to evaluate activities of phenol sulfo- and glucuronyl transferases. Structural similarities between pro-MII and phenolic compounds were the reason

for choosing these substrates for evaluating liver conjugation activities in Trial I. In Trial II, pro-MII was used for the conjugation studies.

The rates of sulfation activity towards 2-naphthol in Trial I were within the range previously reported for pigs (Smith et al., 1984). In both trials, sulfation activity was higher in pigs with low skatole levels in the fat and it was negatively correlated with fat skatole levels in Trial II. This indicates a relationship between the rapid metabolic clearance of skatole and sulfation. In particular, the sulfation of pro-MII may be important as this was the only skatole metabolite that formed a sulfoconjugate in this study. Thus, it is probably the sulfation of pro-MII that results in a rapid metabolic clearance of skatole. Synthesis of pro-MII must be important also, and a tendency for increased synthesis of pro-MII in samples with low skatole levels was observed ($P = .073$).

Although sulfation appears to be the major conjugation reaction involved in skatole metabolism, glucuronidation is also involved. Pro-MII has been found in pigs not only as a sulfate but also as a glucuronyl conjugate (Bæk et al., 1995) and it was easily glucuronidated in this study. Glucuronidation and sulfation are known as competing reactions (Tamellini et al., 1991b), so whether a substrate is glucuronidated or sulfated can depend on the relative activities of both enzymes. Such a competition towards pro-MII was suggested in this study, since a trend for reduced formation of the sulfate conjugate of pro-MII was observed in the assays of skatole metabolites with UDPGA ($P = .103$). UDP-glucuronyl transferases are a group of proteins that are very closely related structurally, but are functionally heterogeneous (Tephly and Burchell, 1990). This heterogeneity was observed in Trial I as p-nitrophenol glucuronidation did not differ between the treatments while 2-naphthol glucuronidation did. The rates of p-nitrophenol glucuronidation corresponded to those reported for pigs previously (Smith at al., 1984). 2-Naphthol formed a conjugate with glucuronyl faster in pigs with high than with low fat skatole levels and glucuronidation was positively correlated with fat skatole levels. A

similar relationship was indicated in Trial II. The pigs with high skatole levels (L) tended to have a higher rate of glucuronidation of pro-MII than the other treatments (P = .069). If it is the sulfation of pro-MII that is required for effective skatole clearance, formation of the glucuronyl conjugate of pro-MII may have an opposite effect to sulfation on fat skatole levels. In both trials, the ratios of glucuronidation:sulfation were higher in pigs with high skatole levels. The ratios were more closely correlated with sulfation than with glucuronidation activities indicating that it is the sulfotransferase activities that vary the most and determine whether pro-MII is sulfated or glucuronidated. Additionally, sulfation but not glucuronidation of pro-MII was correlated with pro-MII formation, P450IIE1 and negatively correlated fat skatole levels in Trial 2. Therefore, glucuronidation of the skatole metabolite F-1 may not be important in the metabolic clearance of skatole although F-1 synthesis seems to have a positive effect. Confirmation of the role of F-1 in the clearance of skatole would require further study.

Pro-MII was also degraded when no cofactors were added to the microsomal incubations. The differences in pro-MII degradation were seen among pigs with different levels of P450IIE1 in liver and skatole in fat and were correlated with several microsomal skatole metabolites. This suggests that pro-MII degradation might be due to the action of enzymes involved in skatole metabolism. It appears that pro-MII could be partially metabolized and partially conjugated. Both transformations could be important in the clearance of skatole, since higher rates of pro-MII degradation were found in the low skatole group of pigs.

These results indicate that both oxidation and conjugation reactions are involved in skatole metabolism and affect the levels of skatole in the fat. The expression of pig liver cytochrome P450IIE1, responsible for the oxidative skatole metabolism, is probably genetically regulated, as it has been indicated in humans (Stephens et al., 1994). It may also be dependent on sex steroid hormones, since the levels of cytochrome P450IIE1 in the liver were high in all female pigs

but varied substantially among intact male pigs (Squires and Lundström, 1997).

Genetic polymorphisms and a high degree of inheritance have been reported for both sulfotransferases (Falany, 1991; Nonneman et al., 1995) and UDP glucuronyl-transferases (Matsui and Watanabe,1982; Kroemer and Klotz 1992) in several mammalian species. The activities of transferases are also under control of sex steroids. The glucuronidation of p-nitrophenol and estrogens was higher in male than female rats (Zhu et al., 1995) confirming early reports of the stimulating effect of testosterone and inhibitory effect of estrogens on glucuronidation activities in rodents (Dutton, 1966). Estrogens increase while androgens decrease hydroxysteroid sulfotransferase activity, while the opposite effects are seen on phenol sulfotransferase in rats (Matsui and Homma, 1994). Sexual differences in the activities of sulfotransferases have been frequently observed in laboratory animals. In contrast, no pronounced differences in glucuronidation or sulfation activities between sexes were observed in humans (Tamellini et al., 1991a). Sulfotransferases are regulated by pituitary growth hormone and thyroid hormone (Matsui and Homma, 1994).

The regulation of both phase I and phase II metabolism of skatole by sex hormones may be the explanation why high fat skatole levels are found mostly in intact male pigs but not in gilts or castrates. The relationship between high levels of skatole and androgens, androstenone and glucocorticoids, (Claus et al., 1994) in male pigs is with agreement with this mechanism for the regulation of fat skatole levels. On the other hand, skatole levels were positively correlated with estrogens but not testosterone (Squires and Lundström, 1997). The exact role of sex hormones needs to be clarified since pigs, as opposed to males of most other species, express high levels of both estrogens and androgens, and both hormones have a synergistic effect on sexual development of male pigs (Joshi and Reaside, 1973).

While the present invention has been described with reference to what are presently considered to be the preferred examples,

it is to be understood that the invention is not limited to the disclosed examples. To the contrary, the invention is intended to cover various modifications and equivalent arrangements included within the spirit and scope of the appended claims.

All publications, patents and patent applications are herein incorporated by reference in their entirety to the same extent as if each individual publication, patent or patent application was specifically and individually indicated to be incorporated by reference in its entirety.

Table 1. Formation of MII and MIII and the disappearance of skatole in microsomal incubations with different cytochrome P450 inhibitors.

			Metabolite[a]		
	Inhibitor	**Concentration, mM**	**pro-MII**	**MIII**	**Skatole**
	SKF 252A	.025	59.5*	101.2	107.0
		.05	49.7*	73.6*	104.6
		.1	38.7*	64.3*	106.1
		.2	38.2*	57.1*	115.3
	P-value[b]		.013	.005	.692
	SE[c]		4.7	4.6	6.6
	Metyrapone	.025	25.8*	ND[d]	105.5
		.05	14.7*	ND	115.8*
		.1	11.6*	ND	129.4*
	P-value		.001	ND	.008
	SE		4.2	ND	3.9
	Chlorzoxazone	.025	82.1*	106.5*	102.9
		.05	74.4*	102.6	105.6
		.1	19.6*	62.7*	113.2*
		.2	9.7*	56.8*	126.7*
	P-value		.001	.001	.007
	SE		2.8	1.81	3.82
	DAS	.025	49.6*	80.2	112.3*
		.05	40.4*	52.3*	118.8*
		.1	35.6*	44.2*	132.1*
		.2	30.9*	30.7*	133.4*
	P-value		.001	.001	.001
	SE		3.7	6.1	1.2

[a]Values are expressed as the least-squares means (n = 3) % of the peak area of control incubations.

[b]Probability value for the differences among concentrations, including control.

[c]Standard error of least-squares mean.

[d]Not determined.

*Values different from control (P < 05).

Table 2. Rates of skatole metabolism and activities of UDP-glucuronyltransferase and sulfotransferase in the liver of intact males with high and low levels of skatole in fat, Trial 1

Item	High (n = 9)	Low (n = 9)	SE[a]	P - value
Level of skatole in fat, ppm[b]	.42	.06	.06	.001
Rate of skatole metabolism, nmol/mg protein/min[b]	.325	.128	.55	.023
Rate of p-nitrophenol glucuronide formation, nmol/mg protein/min	8.47	7.53	.62	.307
Rate of 2-naphthol glucuronide formation, nmol/mg protein/min	29.5	22.1	1.9	.012
Rate of 2-naphthol sulfation, nmol/mg protein/minc.	068	.155	.03	.049
Ratio of 2-naphthol glucuronide formation:2-naphthol sulfation[b]	5062	211	3034	.026

[a] Standard error of the least-squares mean.
[b]Statistical comparisons based on data transformed to common logarithms.
[c]Statistical comparisons based on data transformed to square root values.

Table 3. Levels of skatole in fat and the formation of skatole metabolitesa in liver microsomes from intact male pigs with different levels of cytochrome P450IIE1 in the liver, Trial 2 (Least-squares means ± standard errors)

Item	Low (n = 7)	Medium (n = 8)	High (n = 7)	P - value
P450IIE1 in liver[b]	11.7e ± 5.2	53.0[d] ± 4.8	99.5[c] ± 5.2	.001
Skatole in fat, ppm[f]	.27c ± .03	.13[d] ± .03	.09[d] ± .03	.001
pro-MII [f]	9.9 ± 5.1	28.7 ± 4.7	21.4 ± 5.1	.073
F-1	126[d] ± 46	352[c] ± 43	278[c] ± 46	.007
F-2[f]	126 ± 20.4	35.6 ± 19.1	36.2 ± 20.4	.327
F-3[f]	144 ± 24	70 ± 23	103 ± 24	.087
MIII[f]	20.2 ± 2.7	20.5 ± 2.5	17.7 ± 2.7	.618
UV-1[f]	30.6[c] ± 3.2	16.8[d] ± 3.0	18.0[d] ± 3.2	.023
UV-3	43.4 ± 4.8	27.9 ± 4.4	34.1 ± 4.8	.084
Ratio pro-MII:MIII[f]	.48 ± .38	1.58 ± .35	1.55 ± .38	.141
Skatole metabolism, nmol/mg protein/min	.198 ± .024	.156 ± .22	.147 ± .024	.311
pro-MII sulfation[f]	201[d] ± 375	936c ± 351	1172[c] ± 375	.004
pro-MII glucuronidation	1347 ± 1396	873 ± 1305	1072 ± 1396	.069
Ratio pro-MII gluc:sulf[f]	19.5[c] ± 2.9	1.9[d] ± 2.7	3.1[d] ± 2.9	.003
pro-MII degradation[f]	258[c] ± 29.6	127[d] ± 27.7	117[d] ± 29.6	.013

[a]Concentrations of skatole metabolites expressed in arbitrary units of peak area derived from HPLC analysis.

[b]Concentrations expressed in arbitrary units derived from Western blotting analysis (Squires and Lundstr^m, 1997).
[cde]Means with different superscripts within a raw differ ($P < .05$).
[f]Statistical comparisons based on data transformed to common logarithms.

Table 4. Correlations between the levels of skatole in the fat, cytochrome P450IIE1 in the liver, oxidation and conjugation metabolitesa of skatole in microsomal incubations, Trial 2.

			Oxidative metabolites of skatole							Conjugation of pro-MII				
	P450IIE1 in liver[b]	Skatole in fat	pro-MII	F-1	F-2	F-3	MIII	UV-1	UV-3	Ratio pro-MII: MIII	Skatole metab-olism	Sulfation	Glucu-ronida-tion	Ratio gluc:sulf.
Skatole in fat, ppm[c]	-.79***													
pro-MII[c]	.33 NS	-.35 NS												
F-1	.41 NS	-.59**	.73***											
F-2[c]	-.16 NS	.07 NS	.02 NS	-.23 NS										
F-3[c]	-.19 NS	.15 NS	-.35 NS	-.46*	.60**									
MIII[c]	-.26 NS	.02 NS	-.19 NS	-.08 NS	.37 NS	.63**								
UV-1[c]	-.39 NS	.27 NS	-.38 NS	-.42 NS	.66***	.76***	.60**							
UV-3	-.25 NS	.16 NS	-.49*	-.48*	.67***	.87***	.68***	.88***						
Ratio pro-MII:MIII[c]	.38 NS	-.32 NS	.94***	.67***	-.10 NS	-.52*	-.50*	-54**	-.67***					
Skatole metabolism nmol/mg prot./min	-.28 NS	.22 NS	-.56**	-.45*	.15 NS	.70***	.58**	.56**	.70***	-.68***				
pro-MII Sulfation[c]	.57**	-.56**	.70***	.62**	-.13 NS	-.45*	-.32 NS	-.53*	-.54**	.72***	.54**			
pro-MII Glucuronidation	-.34 NS	.35 NS	-.35 NS	-.24 NS	.27 NS	.40 NS	.17 NS	.47*	.43*	-.37 NS	-.17 NS	.45*		
Ratio pro-MII gluc.:sulf.[c]	-.55**	.56**	-.70***	-.58**	.19 NS	-.52*	.33 NS	.59**	.59**	-.73***	-.53*	-.97***	.67***	
pro-MII Degradation[c]	-.51*	.42 NS	-.18 NS	-.36 NS	.37 NS	.57**	.43*	.69***	.59**	-.30 NS	-.49*	-.34 NS	.13 NS	.48*

*P < .05; **P < .01; ***P < .001; NS: not significant (P > .05).

[a]Concentrations of skatole metabolites expressed in arbitrary units of peak area derived from HPLC analysis.

[b]Concentrations expressed in arbitrary units derived from Western blotting analysis (Squires and Lundstr^m, 1997).

[c]Correlations based on data transformed to common logarithms.

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THE EMBODIMENTS OF THE INVENTION IN WHICH AN EXCLUSIVE PROPERTY OR PRIVILEGE IS CLAIMED ARE DEFINED AS FOLLOWS:

1. A method for reducing or preventing boar taint comprising enhancing the metabolism of skatole in a pig.

2. A method according to claim 1 comprising enhancing the activity of the CYP2E1 enzyme in a pig.

3. A method according to claim 2 wherein the activity of the CYP2E1 enzyme is enhanced by administering

(a) a substance that increases the activity of the CYP2E1 enzyme; or

(b) a substance that induces or increases the expression of the CYP2E1 gene.

4. A method according to claim 2 wherein a nucleic acid sequence encoding a CYP2E1 enzyme is introduced into a pig.

5. A method according to claim 1 comprising enhancing the activity of the sulfotransferase enzyme in a pig.

6. A method according to claim 5 wherein the activity of the sulfotransferase enzyme is enhanced by administering

(a) a substance that increases the activity of the sulfotransferase enzyme; or

(b) a substance that induces or increases the expression of the sulfotransferase gene.

7. A method according to claim 5 wherein a nucleic acid sequence encoding a sulfotransferase enzyme is introduced into a pig.

8. A method according to claim 1 comprising inhibiting the activity of the glucuronyl transferase enzyme in a pig.

9. A method according to claim 8 wherein the activity of the glucuronyl transferase enzyme is decreased by administering

(a) a substance that decreases the activity of the glucuronyl transferase enzyme; or

(b) a substance that reduces or decreases the expression of the glucuronyl transferase gene.

10. A method according to claim 8 wherein an antisense nucleic acid sequence that is complementary to a nucleic acid sequence encoding a glucuronyl transferase enzyme is introduced into a pig.

11. A method of screening for a substance that enhances the activity of CYP2E1 comprising assaying for a substance which selectively (i) enhances CYP2E1 activity, or (ii) enhances transcription and/or translation of the gene encoding CYP2E1.

12. A method is provided for screening for a substance that enhances skatole metabolism in a pig by enhancing CYP2E1 activity comprising the steps of :

(a) reacting a substrate of CYP2E1 and CYP2E1, in the presence of a test substance, under conditions such that CYP2E1 is capable of converting the substrate into a reaction product;

(b) assaying for reaction product, unreacted substrate or unreacted CYP2E1;

(c) comparing to controls to determine if the test substance selectively enhances CYP2E1 activity and thereby is capable of enhancing skatole metabolism in a pig.

13. A method of screening for a substance that enhances the activity of sulfotransferase comprising assaying for a substance which

selectively (i) enhances sulfotransferase activity, or (ii) enhances transcription and/or translation of the gene encoding sulfotransferase.

14. A method for screening a substance that enhances

(a) reacting a substrate of sulfotransferase and sulfotransferase, in the presence of a test substance, under conditions such that sulfotransferase is capable of converting the substrate into a reaction product;

(b) assaying for reaction product, unreacted substrate or unreacted sulfotransferase;

(c) comparing to controls to determine if the test substance selectively enhances sulfotransferase activity and thereby is capable of enhancing skatole metabolism in a pig.

15. A method of screening for a substance that inhibits the activity of glucuronyl transferase comprising assaying for a substance which selectively (i) inhibits glucuronyl transferase activity, or (ii) inhibits transcription and/or translation of the gene encoding glucuronyl transferase.

16. A method for screening a substance that inhibits

(a) reacting a substrate of glucuronyl transferase and glucuronyl transferase, in the presence of a test substance, under conditions such that glucuronyl transferase is capable of converting the substrate into a reaction product;

(b) assaying for reaction product, unreacted substrate or unreacted glucuronyl transferase;

(c) comparing to controls to determine if the test substance selectively inhibits glucuronyl transferase activity and thereby is capable of inhibiting skatole metabolism in a pig.

17. A method for screening for a substance that enhances skatole metabolism by enhancing transcription and/or translation of the gene encoding CYP2E1 comprising the steps of:

(a) culturing a host cell comprising a nucleic acid molecule containing a nuclei acid sequence encoding CYP2E1 and the necessary elements for the transcription or translation of the nucleic acid sequence, and optionally a reporter gene, in the presence of a test substance; and

(b) comparing the level of expression of CYP2E1, or the expression of the protein encoded by the reporter gene with a control cell transfected with a nucleic acid molecule in the absence of the test substance.

18. A method for screening for a substance that enhances skatole metabolism by enhancing transcription and/or translation of the gene encoding sulfotransferase comprising the steps of:

(a) culturing a host cell comprising a nucleic acid molecule containing a nuclei acid sequence encoding sulfotransferase and the necessary elements for the transcription or translation of the nucleic acid sequence, and optionally a reporter gene, in the presence of a test substance; and

(b) comparing the level of expression of sulfotransferase, or the expression of the protein encoded by the reporter gene with a control cell transfected with a nucleic acid molecule in the absence of the test substance.

19. A method for screening for a substance that enhances skatole metabolism by inhibiting transcription and/or translation of the gene encoding glucuronyl transferase comprising the steps of:

(a) culturing a host cell comprising a nucleic acid molecule containing a nuclei acid sequence encoding glucuronyl transferase and the necessary elements for the transcription or

translation of the nucleic acid sequence, and optionally a reporter gene, in the presence of a test substance; and

(b) comparing the level of expression of glucuronyl transferase, or the expression of the protein encoded by the reporter gene with a control cell transfected with a nucleic acid molecule in the absence of the test substance.

20. A composition for reducing or preventing skatole metabolism comprising administering an effective amount of a substance that enhances skatole metabolism.

21. A composition according to claim 20 wherein the substance is selected from the group consisting of

(a) a substance that increases the activity of the CYP2E1 enzyme;

(b) a substance that induces or increases the expression of the CYP2E1 gene;

(c) a substance that increases the activity of the sulfotransferase enzyme;

(d) a substance that induces or increases the expression of the sulfotransferase gene;

(e) a substance that decreases the activity of the glucuronyl transferase enzyme; and

(f) a substance that reduces or decreases the expression of the glucuronyl transferase gene.

22. A composition according to claim 20 wherein the substance is identified according to the method of any one of claims 11-19.

FIGURE 1A

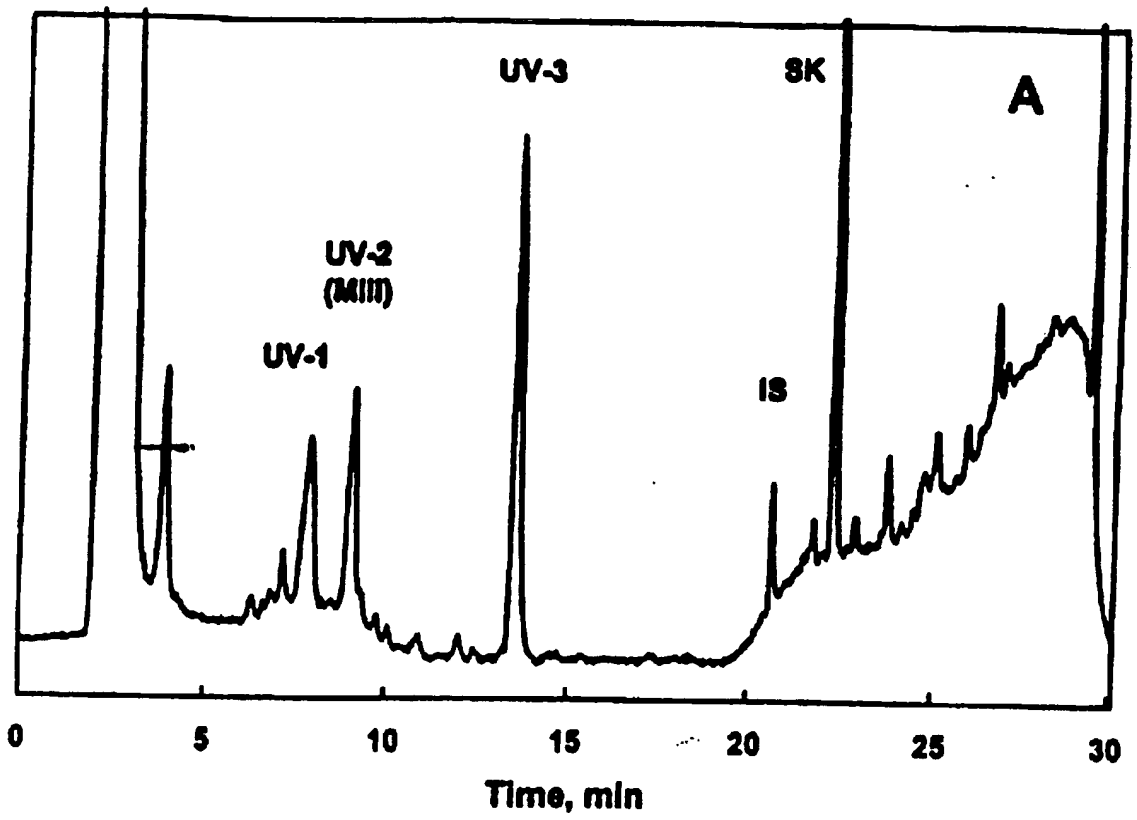

FIGURE 1B

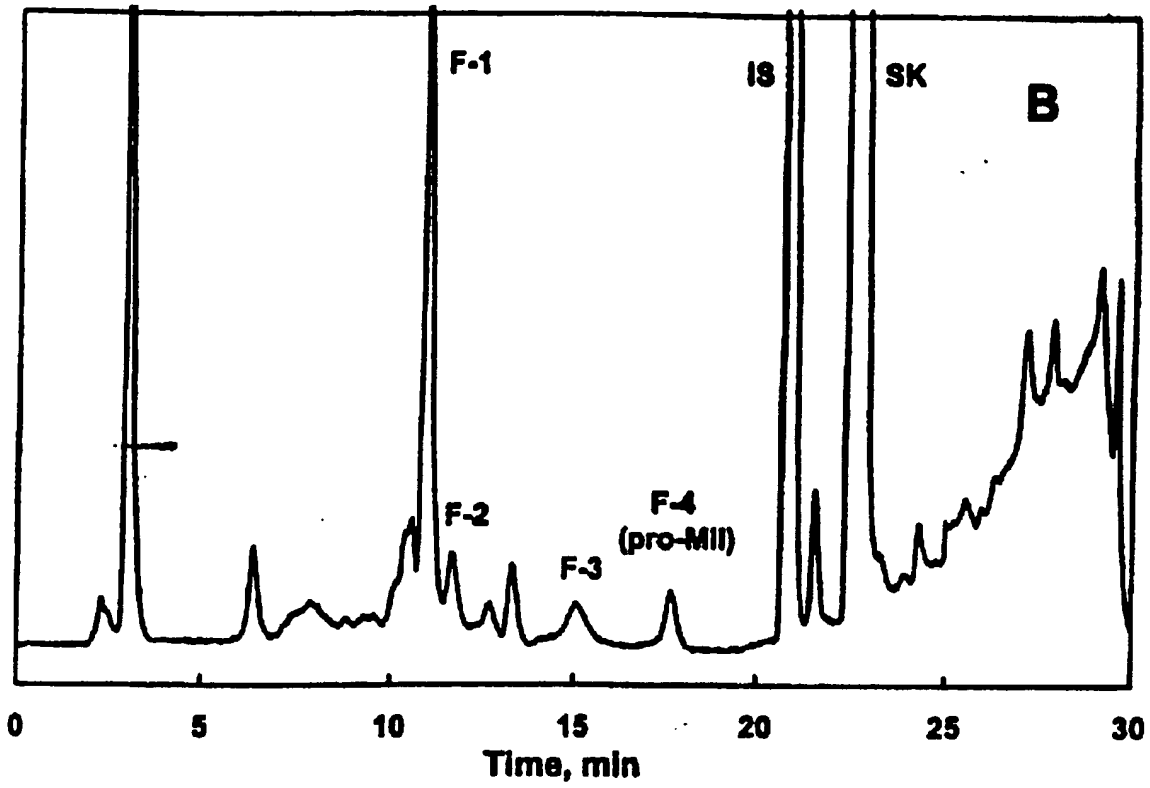

FIGURE 2

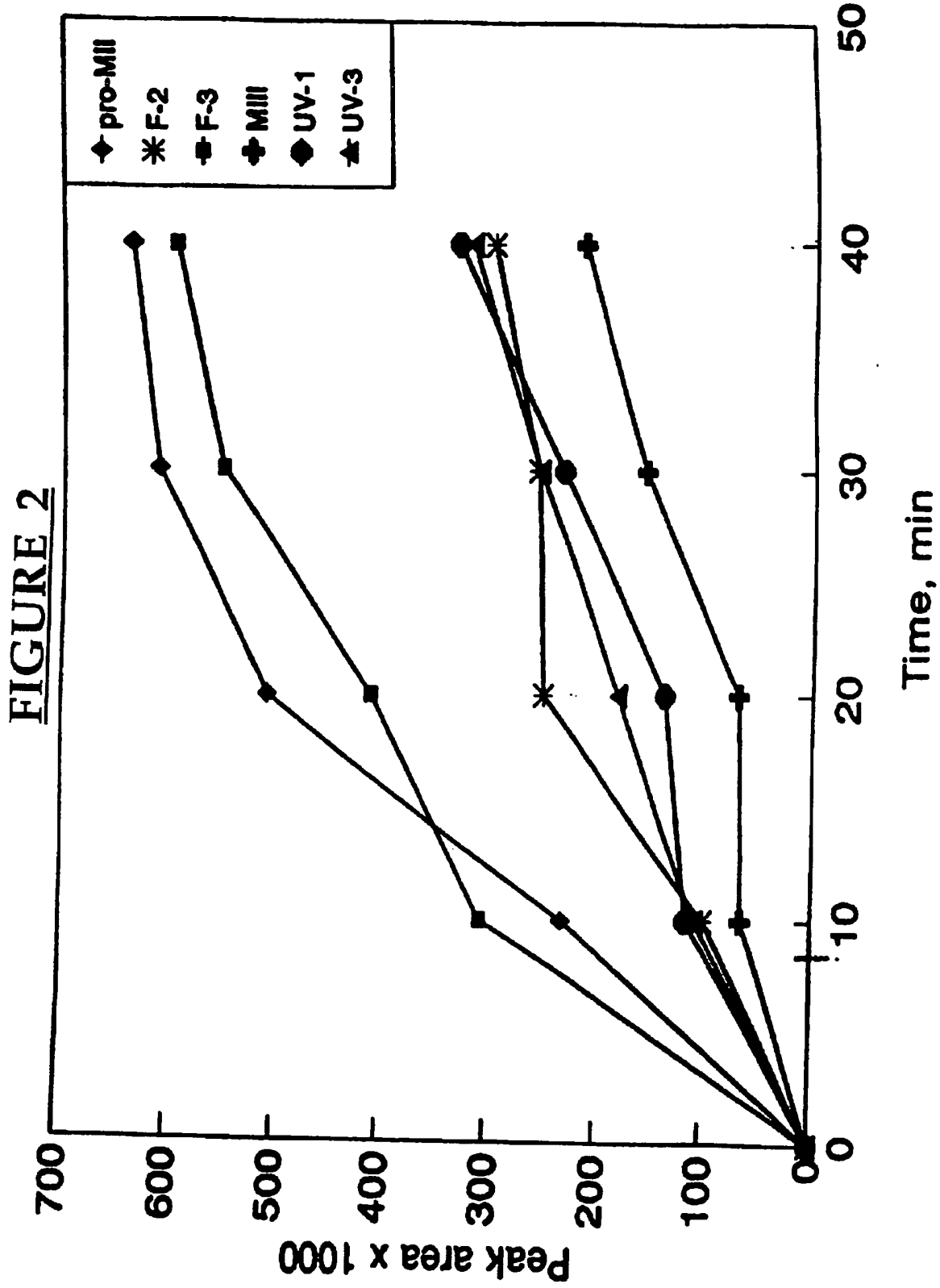
pro-MIII
F-2
F-3
MIII
UV-1
UV-3
Peak area x 1000
Time, min
0
100
200
300
400
500
600
700
0
10
20
30
40
50

FIGURE 3

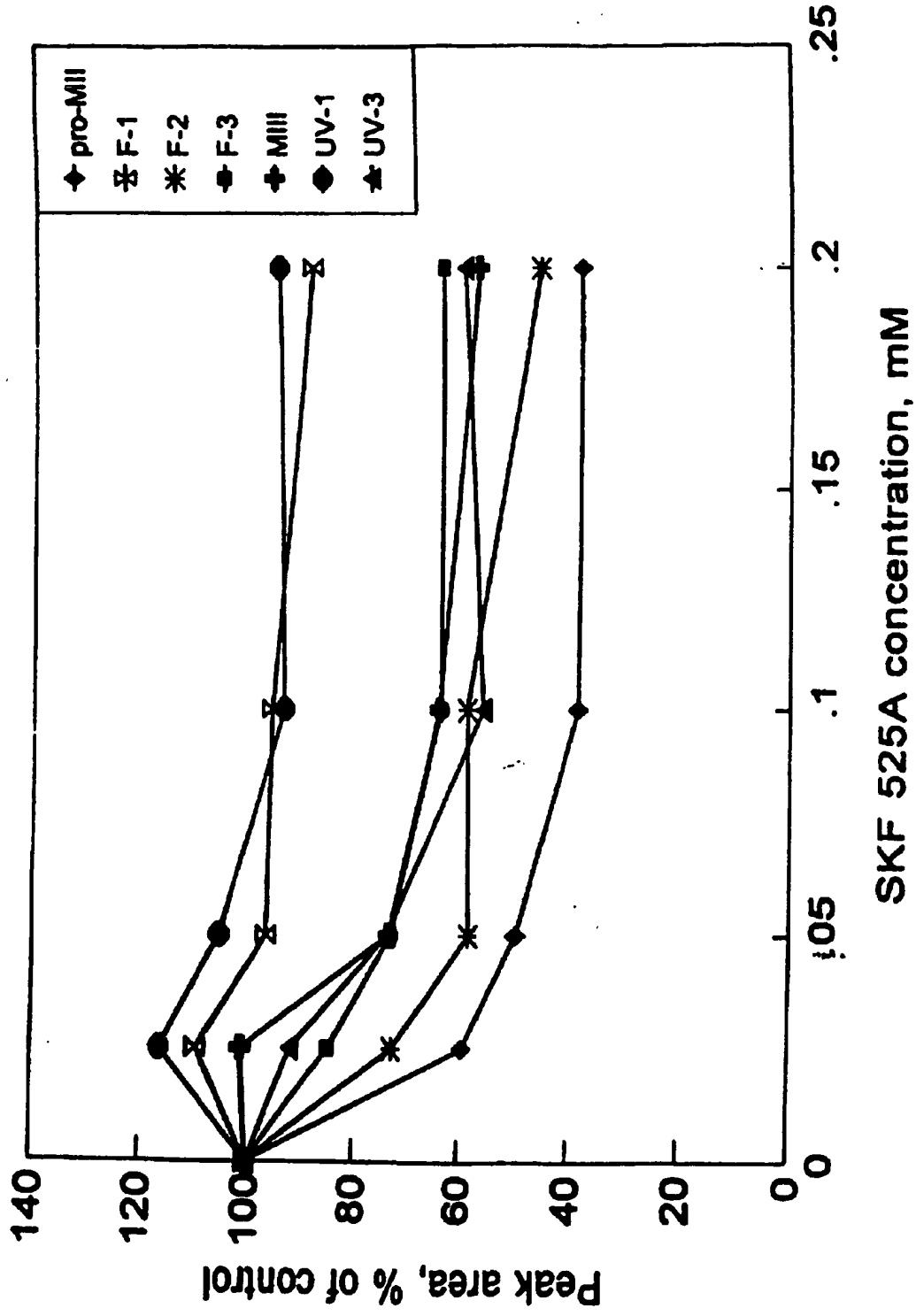
pro-MII
F-1
F-2
F-3
MIII
UV-1
UV-3
Peak area, % of control
140
120
100
80
60
40
20
0
0
.05
.1
.15
.2
.25
SKF 525A concentration, mM

FIGURE 4

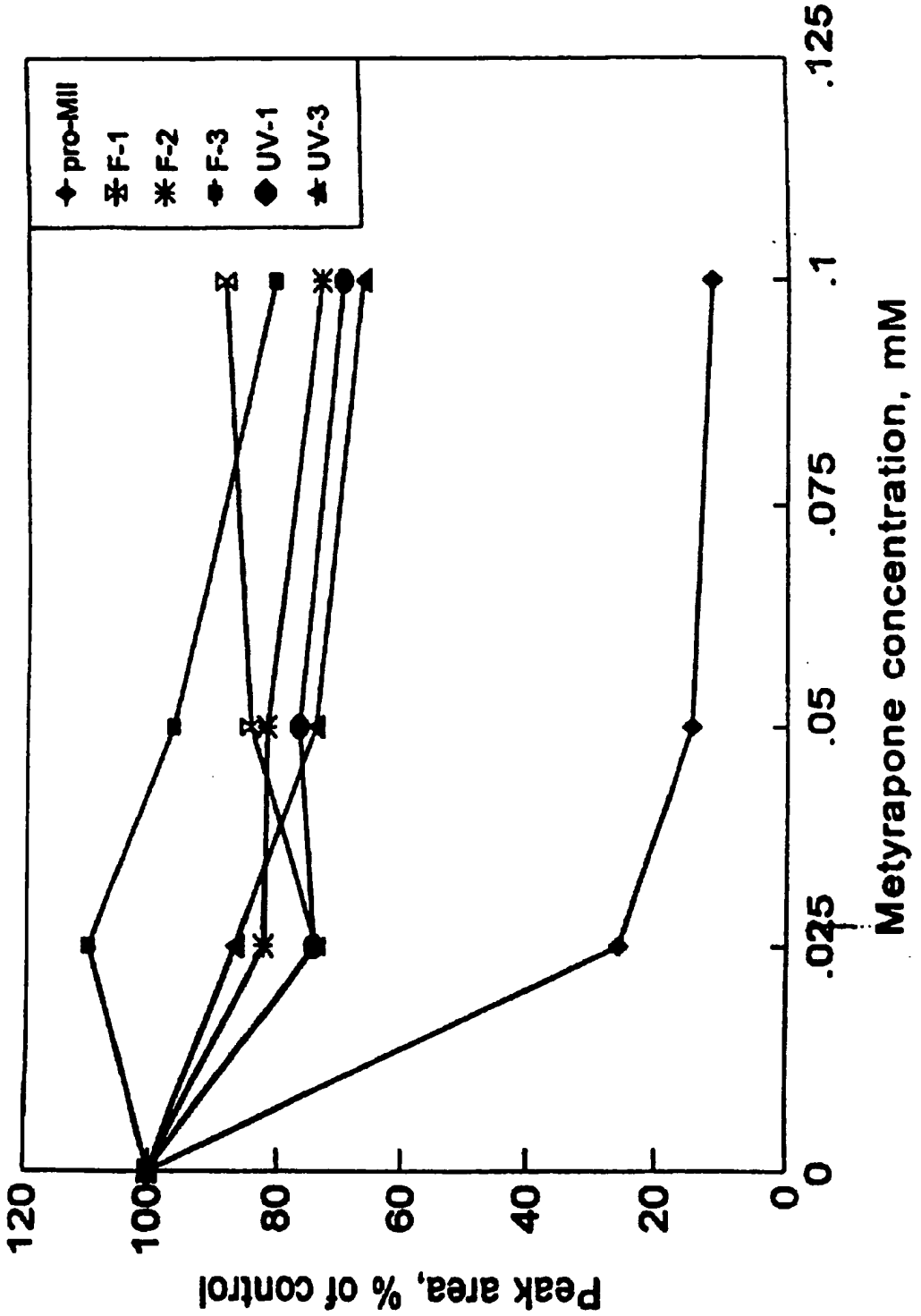

FIGURE 5

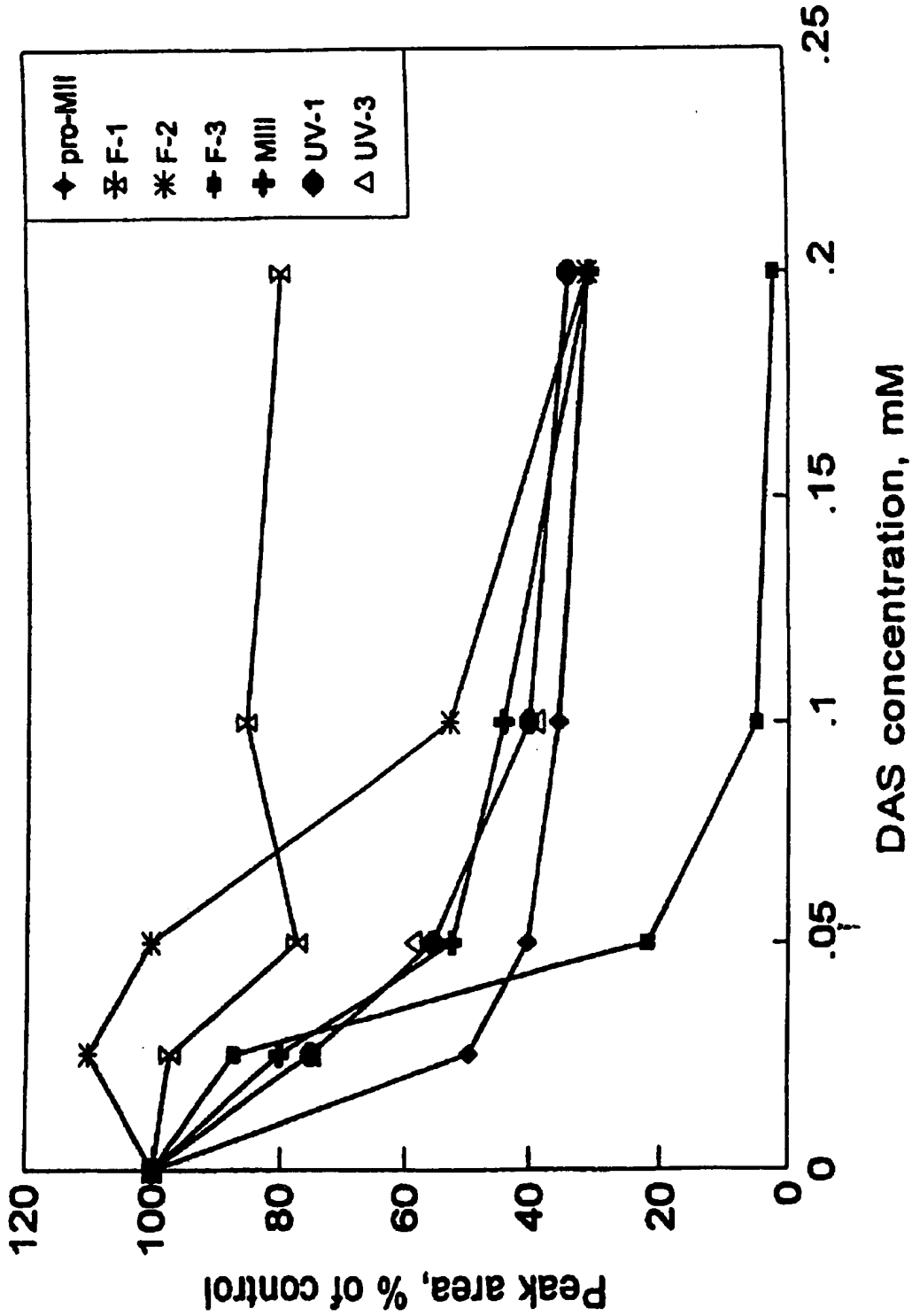
pro-MII
F-1
F-2
F-3
MIII
UV-1
UV-3
Peak area, % of control
120
100
80
60
40
20
0
0
.05
.1
.15
.2
.25
DAS concentration, mM

FIGURE 6

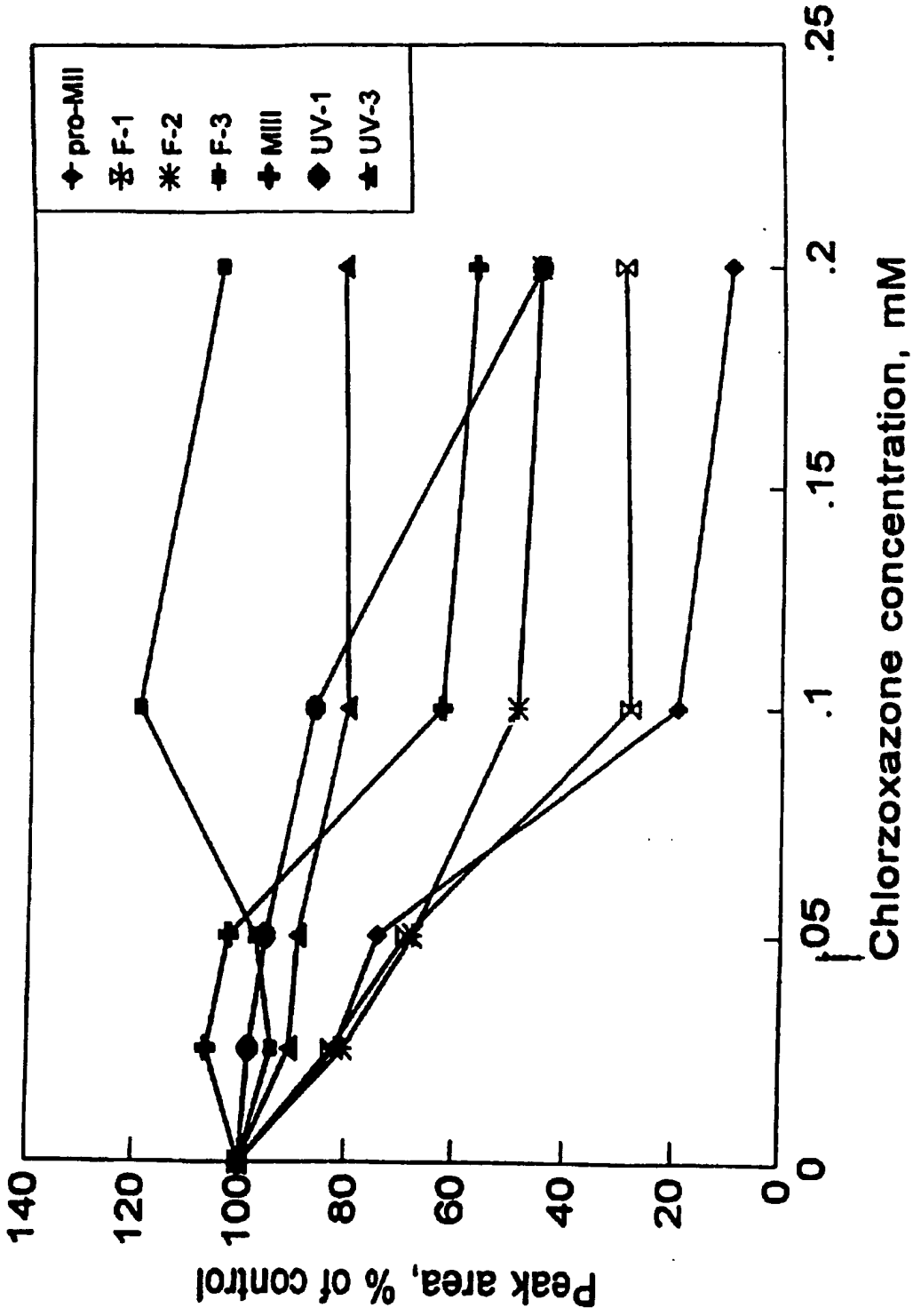
pro-MII
F-1
F-2
F-3
MIII
UV-1
UV-3
Peak area, % of control
140
120
100
80
60
40
20
0
0
.05
.1
.15
.2
.25
Chlorzoxazone concentration, mM

FIGURE 7

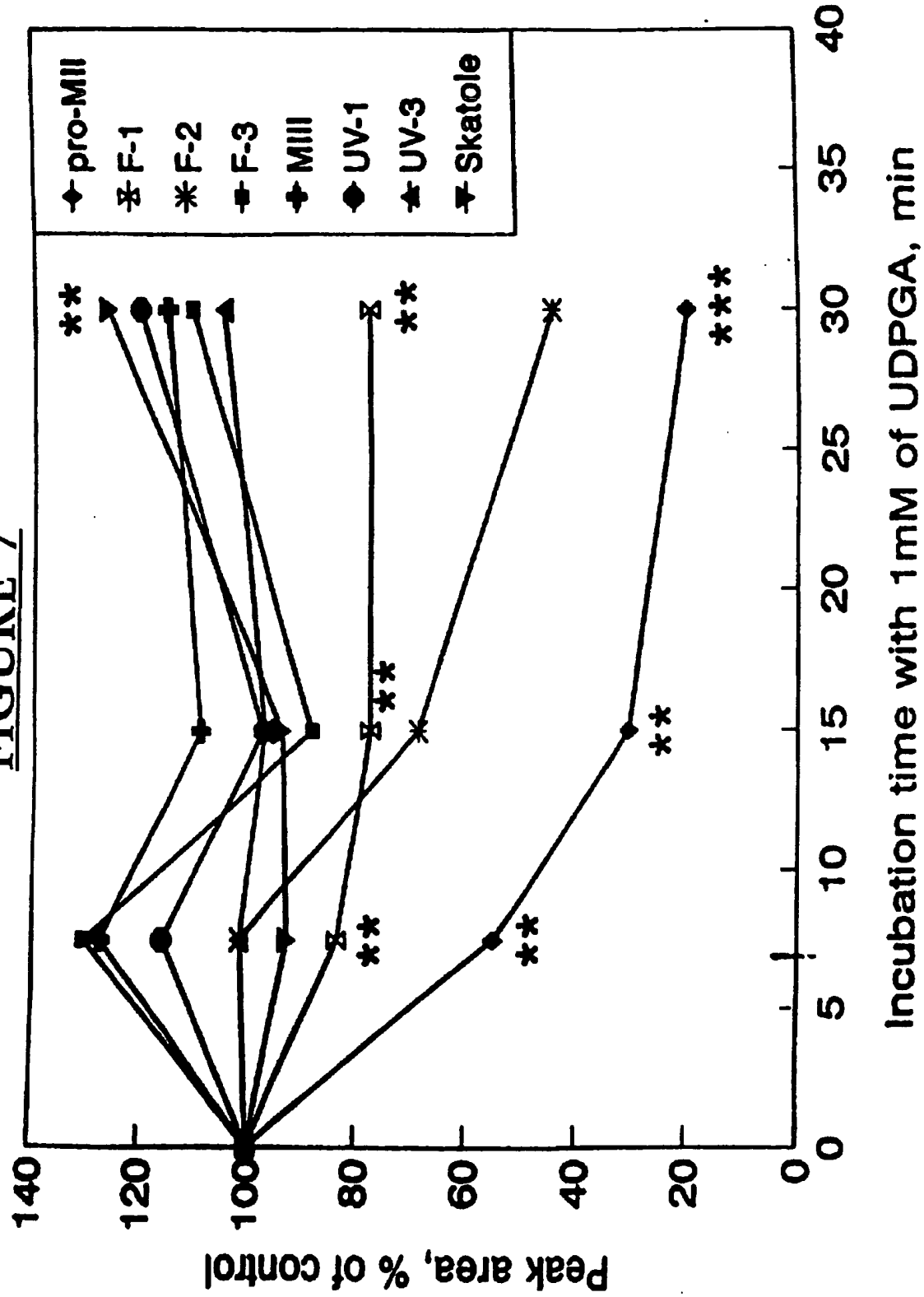
pro-MII
F-1
F-2
F-3
MIII
UV-1
UV-3
Skatole
Peak area, % of control
Incubation time with 1mM of UDPGA, min
0
20
40
60
80
100
120
140
5
10
15
20
25
30
35
40
**
***

FIGURE 8

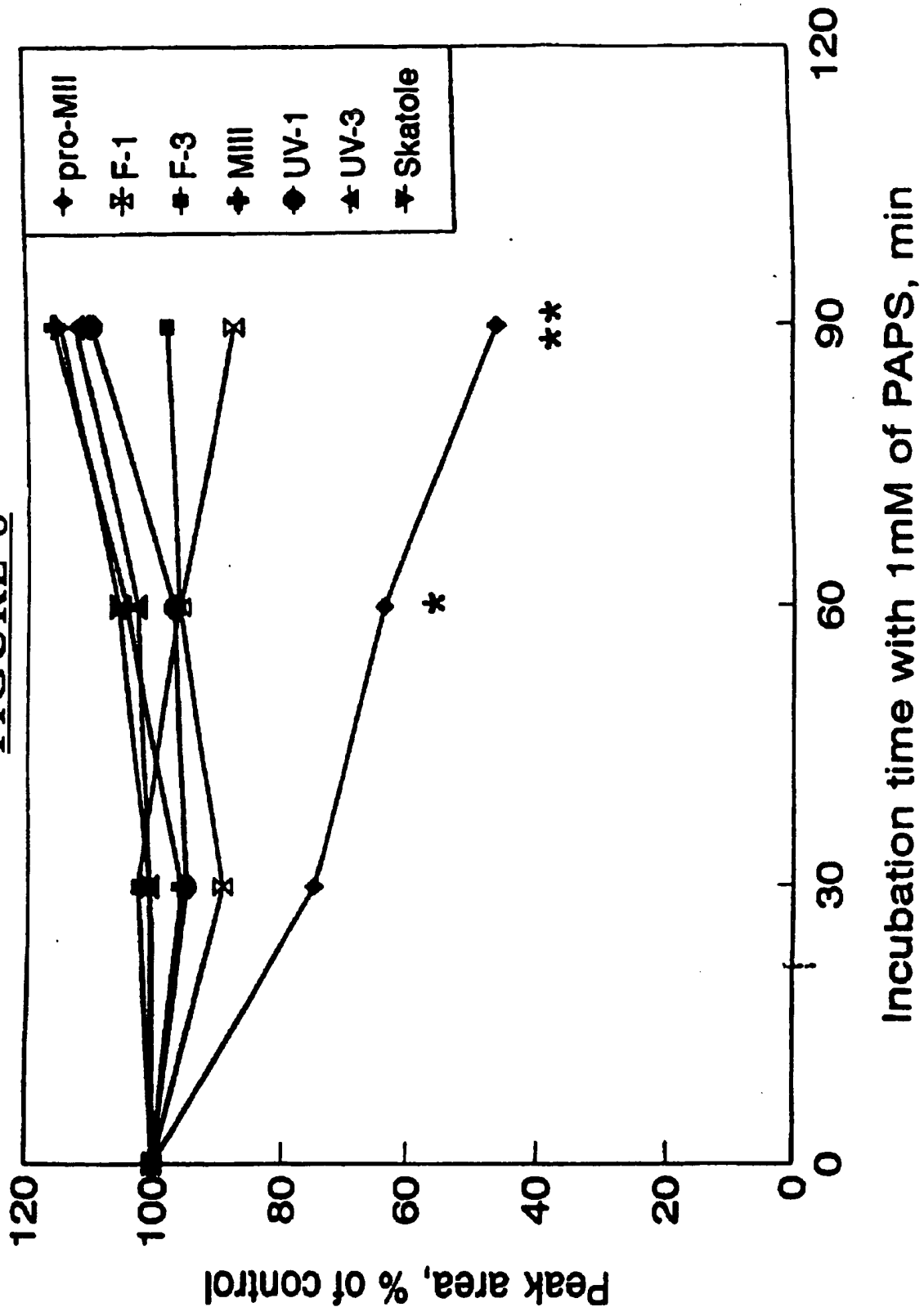
pro-MII
F-1
F-3
MIII
UV-1
UV-3
Skatole
120
100
80
60
40
20
0
Peak area, % of control
0
30
60
90
120
Incubation time with 1mM of PAPS, min
*
**

FIGURE 9